

Coagulase Mannitol HiVeg™ Broth Base

MV277

Intended use

With plasma it is recommended for simultaneous detection of coagulase production and mannitol fermentation in the differentiation of Staphylococci

Composition**

Ingredients	g / L
HiVeg™ infusion	10.000
HiVeg™ peptone	10.000
D-Mannitol	10.000
Sodium chloride	5.000
Phenol red	0.025
Final pH (at 25°C)	7.4±0.2

**Formula adjusted, standardized to suit performance parameters

Directions

Suspend 35.0 grams in 1000 ml purified/ distilled water. Heat if necessary to dissolve the medium completely. Sterilize by autoclaving at 118-121°C for 15 minutes. Cool to 45-50°C. Just before use, add 7-15% v/v sterile, pre-tested, rabbit plasma (coagulase plasma) to the basal medium. Mix well and dispense into sterile tubes.

Principle And Interpretation

Coagulase Mannitol HiVeg™ Broth Base is used for the isolation of *Staphylococcus aureus* from clinical specimens and for differentiation of *Staphylococcus aureus* from other species on the basis of coagulase production and mannitol fermentation. Chapman for the first time introduced medium for selective isolation and differentiation of Staphylococci (1). Tellurite-glycine media were designed by Zebovitz et al (2) and Marwin (3) for selectively isolating coagulase positive Staphylococcal species. Present medium is based on Esber and Faulconer formulation (4). Coagulase Mannitol HiVeg™ Broth Base is prepared by using vegetable peptones in place of animal based peptones which make the media free of BSE/ TSE risks. Coagulase Mannitol HiVeg™ Broth Base simultaneously detects mannitol fermentation and coagulase production by Staphylococci (5). Coagulase Mannitol Broth Base is a good substrate for Staphylococci as well as other fastidious bacteria. Coagulase production and mannitol fermentation observed in Coagulase Mannitol HiVeg™ Broth Base is presumptive identification of pathogenic Staphylococci (6). Coagulase production is dependent on the presence of a fermentable sugar like mannitol in this case. It is also dependent on the presence of a protein factor in the HiVeg™ infusion and blood plasma (4). When mannitol is fermented, the pH of the medium drops. This drop in pH is indicated by the change in colour of the phenol red, which turns yellow, and exhibit yellow medium. An opaque broth due to coagulated plasma forms due to growth of coagulase positive organisms.

Staphylococcus epidermidis a coagulase negative and mannitol non-fermenting species, does not change the colour of the medium. Coagulase negative species may ferment mannitol and produce a yellow colour but opacity will not be formed. Production of gas can be determined by placing a small inverted Durham's tube in the medium tube. Mutant or old cultures of *Staphylococcus aureus* may be weak coagulase producers. They should be freshly subcultured and rechecked. *Escherichia coli* ferments mannitol and may be weakly coagulase positive. For the test, use this medium in 2 to 5 ml amounts after adding about 12-15% plasma. Inoculate by adding about 2 drops of test organism and incubate at 37°C and examine after 2-3 hours and also after 4-5 hours of incubation. Un-inoculated control tubes should also be run in parallel with the fermentation tests.

Type of specimen

Food and dairy samples

Specimen Collection and Handling:

For food and dairy samples, follow appropriate techniques for sample collection and processing as per guidelines (1,8,7). After use, contaminated materials must be sterilized by autoclaving before discarding.

Warning and Precautions :

Read the label before opening the container. Wear protective gloves/protective clothing/eye protection/ face protection. Follow good microbiological lab practices while handling specimens and culture. Standard precautions as per established guidelines should be followed while handling specimens. Safety guidelines may be referred in individual safety data sheets.

Limitations :

1. This medium is general purpose medium and may not support the growth of fastidious organisms.

Performance and Evaluation

Performance of the medium is expected when used as per the direction on the label within the expiry period when stored at recommended temperature.

Quality Control

Appearance

Light yellow to light pink homogeneous free flowing powder

Colour and Clarity of prepared medium

Red coloured, clear to slightly opalescent solution in tubes

Reaction

Reaction of 3.5% w/v aqueous solution at 25°C. pH : 7.4±0.2

pH

7.20-7.60

Cultural Response

Cultural characteristics observed with added 7-15% v/v sterile pretested coagulase plasma after an incubation at 35-37°C for 18-48 hours.

Organism	Inoculum (CFU)	Growth	Acid from Mannitol	Coagulase activity
<i>Staphylococcus aureus</i> subsp. <i>aureus</i> ATCC 25923 (00034*)	50-100	luxuriant	positive reaction, yellow colour	positive reaction, clot formation
<i>Staphylococcus epidermidis</i> ATCC 12228	50-100	luxuriant	negative reaction, no colour change	negative reaction, no clot formation

Key : *Corresponding WDCM numbers.

Storage and Shelf Life

Store between 10-30°C in a tightly closed container and the prepared medium at 2-8°C. Use before expiry date on the label. On opening, product should be properly stored dry, after tightly capping the bottle in order to prevent lump formation due to the hygroscopic nature of the product. Improper storage of the product may lead to lump formation. Store in dry ventilated area protected from extremes of temperature and sources of ignition. Seal the container tightly after use. Product performance is best if used within stated expiry period.

Disposal

User must ensure safe disposal by autoclaving and/or incineration of used or unusable preparations of this product. Follow established laboratory procedures in disposing of infectious materials and material that comes into contact with sample must be decontaminated and disposed of in accordance with current laboratory techniques (7,8).

Reference

1. Chapman, 1946, J. Bact., 51:409.
2. Zebovitz, Evans and Nivens, 1955, J. Bact., 70:686.
3. Marwin, 1958, Am. J. Clin. Pathol., 30:470.
4. Esber and Faulconer, 1959, Am. J. Clin. Pathol., 32:192.
5. Schaub and Merrit, 1960, Bull. Johns Hopkins Hosp., 106:25.
6. Mincheu and Cluff, 1961, J. Chron. Dis., 13:354.
7. Jorgensen, J.H., Pfaller, M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
8. Isenberg, H.D. Clinical Microbiology Procedures Handbook 2nd Edition.

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