



HiCulture™ Transport Swab w/ SCDM w/ Lecithin, Polysorbate MS5335GT 80 & Normal Saline Tube (γ-irradiated)(Triple Pack)

Intended use

Recommended for transporting microbial specimens.

Composition**

Ingredients Tube 1	Gms / Litre
Tryptone	15.000
Soya Peptone	5.000
Sodium chloride	5.000
Lecithin	0.700
Polysorbate 80 (Tween 80)	5.000
Agar	15.000
Final pH (at 25°C)	7.3±0.2

Ingredients Tube 2	Gms / Litre
Sodium chloride	8.500

**Formula adjusted, standardized to suit performance parameters

Directions

1. Identify surface, area or site to be sampled. A sterile square sampling template is recommended when sampling flat surfaces.
2. Open tube containing saline.
3. Insert the flock tip swab into the saline tube.
4. Place the template at the sampling surface & swab in bidirection, in order to achieve maximum uptake of surface material.
5. When sampling is completed, insert the swab into second tube with Soyabean Casein Digest Medium w/ lecithin and Polysorbate 80. Make sure the cap is properly tightened
6. Turbidity in the medium indicates growth.

Principle And Interpretation

Tryptone Soya Agar with Lecithin and Polysorbate 80 is used in RODAC (Replicate Organism Detection and Counting) plates (3) for the detection and enumeration of microorganisms present on surfaces of sanitary importances (6,8).

Tryptone and Soya peptone provide nitrogenous compounds and other nutrients essential for microbial replication. Lecithin and polysorbate 80 (Tween 80) are neutralizers reported to inactivate residual disinfectants from where the sample is collected (1). Lecithin neutralizes quaternary ammonium compounds and polysorbate 80 neutralizes phenolic disinfectants, hexachlorophene, formalin and with lecithin ethanol (2).

Collection of samples from areas before and after the treatment with disinfectant evaluates cleaning procedures in environmental sanitation. The presence and number of microorganisms is determined by the appearance of colonies on the agar surface (7). After counting the colonies, carry out biochemical testing for identification.

Type of specimen

Environmental monitoring

Specimen Collection and Handling

After use, contaminated materials must be sterilized by autoclaving before discarding.

Warning and Precautions

Read the label before opening the container. Wear protective gloves/protective clothing/eye protection/ face protection. Follow good microbiological lab practices while handling specimens and culture. Standard precautions as per established guidelines should be followed while handling specimens. Safety guidelines may be referred in individual safety data sheets.

Limitations

1. Each lot of the medium has been tested for organisms specified on the COA. It is recommended to validate the media for

organisms other than specified as per users requirement.

Performance and Evaluation

Performance of the medium is expected when used as per the direction on the label within the expiry period when stored at recommended temperature.

Quality Control

Appearance

Each pack contains two tubes Sterile SCDM w/ Lecithin & Polysorbate 80 in one tube (A), & Normal Saline (B) in other tube along with two sterile flocced nylon swabs with breakpoint.

Colour

Part A: Light yellow clear liquid, Part B: Colourless liquid

Quantity of Medium

Part A: 5 ml of medium in tubes Part B: 5 ml of medium in tubes

Dose of Irradiation (Kgy)

25.00 -30.00

Sterility check

Passes release criteria

Cultural Response

Viability of following organisms was established for a period of 18-24 hours for bacterial cultures & 48-72 hours for fungal cultures. Organisms grew luxuriantly when recovered on Tryptone Soya Agar (MH290) and incubated at 35-37°C for 18-72 hours.

Organism	Recovery
<i>Bacillus subtilis subsp. spizizenii</i> ATCC 6633 (00003*)	luxuriant
<i>Staphylococcus aureus subsp. aureus</i> ATCC 2593 (0003*)	luxuriant
<i>Staphylococcus aureus subsp. aureus</i> ATCC 6538 (00032*)	luxuriant
<i>Escherichia coli</i> ATCC 25922 (00013*)	luxuriant
<i>Escherichia coli</i> ATCC 8739 (00012*)	luxuriant
<i>Escherichia coli</i> NCTC 9002	luxuriant
<i>Pseudomonas aeruginosa</i> ATCC 27853 (00025*)	luxuriant
<i>Pseudomonas aeruginosa</i> ATCC 9027 (00026*)	luxuriant
<i>Salmonella</i> Abony NCTC 6017	luxuriant
<i>Micrococcus luteus</i> ATCC 9341	luxuriant
<i>Streptococcus pneumoniae</i> ATCC 6305	luxuriant
<i>Salmonella</i> Typhimurium ATCC 14028 (00031*)	luxuriant
<i>Candida albicans</i> ATCC 10231 (00054*)	luxuriant
<i>Candida albicans</i> ATCC 2091 (00055*)	luxuriant
<i>Aspergillus brasiliensis</i> ATCC 16404 (00053*)	luxuriant

Key : (#) Formerly known as *Aspergillus niger*, (*) Corresponding WDCM numbers

Please refer disclaimer Overleaf.

Storage and Shelf Life

Store between 5-25°C. Use before expiry date on the label.

Disposal

User must ensure safe disposal by autoclaving and/or incineration of used or unusable preparations of this product. Follow established laboratory procedures in disposing of infectious materials and material that comes into contact with sample must be decontaminated and disposed of in accordance with current laboratory techniques (4,5).

Reference

1. Brummer, 1976, Appl. Environ. Microbiol., 32:80.
2. Favero (Chairman), 1967, Biological Contamination Control Committee, a state of the art report., Am. Assoc. for contamination control.
3. Hall and Hartnett, 1964, Public Hlth. Rep., 79:1021.
4. Isenberg, H.D. Clinical Microbiology Procedures Handbook 2nd Edition.
5. Jorgensen, J.H., Pfaller, M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
6. MacFaddin J.F., 1985, Media for Isolation-Cultivation-Identification-Maintenance of Medical Bacteria, Vol. I, Williams and Wilkins, Baltimore.
7. Murray PR, Baron, Pfaller, and Tenover (Eds.), 2003, In Manual of Clinical Microbiology, 8th ed., ASM, Washington, D.C.
8. Richardson (Ed.), 1985, Standard Methods for the Examination of Dairy Products, 15th ed., APHA, Washington, D.C.

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