MB518 HiPurA® 96 Plasmid DNA Purification Kit

Kit Contents

Product Code	Reagents provided	MB518
		1X96 Preps
DS0020	Resuspension Solution (HP1)	35 ml
DS0021	Lysis Solution (HP2)	35 ml
DS0022	Neutralization Solution (HN3)	38 ml
DS0032	Wash Solution (HPB)	58 ml
DS0024	Wash Solution Concentrate (HPE)	40 ml
DS0040	Elution Buffer (ET) [10 mM Tris-Cl, pH 8.5]	24 ml
DS0003	RNase A Solution (20 mg/ml)	175 µl
DBPL-96-01	HiPurA® 96-well DNA Plate	1 no
LWB-96	HiPurA® 96-well Block (2.2 ml)	1 no
LWB-16-96	HiPurA® 96-well Block (1.6 ml)	2 nos
PR11	HiPurA® Silicon Pad for sealing	1 no
PR18	HiPurA® Breathable Film	1 no
DVB-96	HiPurA® 96-well V-Block	1 no

Intended Use

Recommended for isolation of DNA from Plasmid.

Introduction

The HiPurA® 96 Plasmid DNA Purification Kit provides a simple and rapid method for purifying plasmid or cosmid DNA from small volume of bacterial cultures (2.0 ml). This kit is not recommended for the purification of large recombinant constructs such as BACs and PACs. The kit uses the principle of silica binding in a 96-well format that eliminates the need for expensive resins, alcohol precipitation and hazardous organic compounds such as phenol and chloroform. This procedure is based on a modified alkaline lysis procedure specialized for high-throughput DNA applications.

HiPurA® 96 Plasmid DNA Purification Kit

The harvested bacterial culture is lysed and collected by centrifugation after which it is applied on to the silica column for binding of the DNA molecules in the presence of high salt concentration. The adsorbed DNA is washed to remove contaminants, and the pure plasmid DNA is eluted in Elution Buffer. The purified plasmid DNA can be used for immediate use in all molecular biology procedures such as digestion with restriction enzymes, cloning, PCR, transfection, *in vitro* translation, blotting and sequencing.

ISO 9001-2015 CERTIFIED
WHO GMP CERTIFIED
ISO 13485-2016 CERTIFIED

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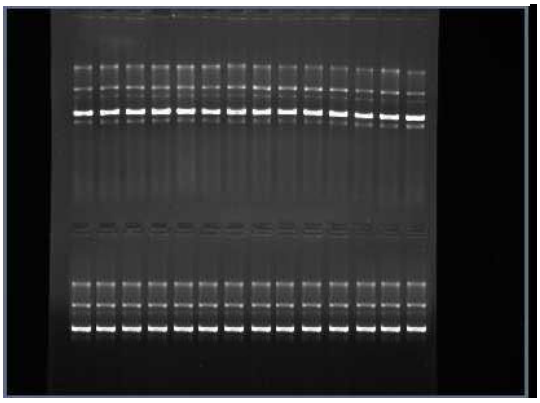
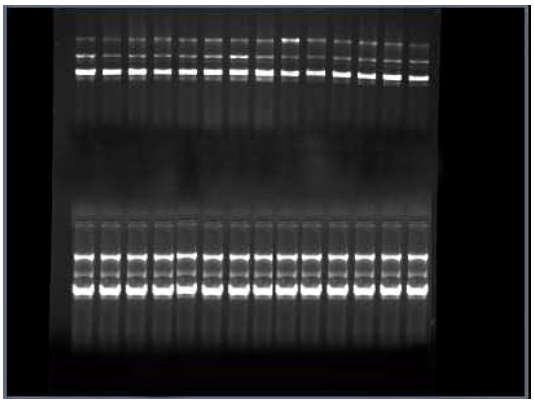
Bacterial culture volume of upto 2.0 ml can be used for plasmid DNA isolation per well. Using enriched media, example 2X YT [YT Agar (Product Code: M1369)], can yield higher plasmid DNA especially in low copy-number vectors, since such media generate higher cell densities for certain vector/host strain combinations than regular LB broth medium (Product Code: M1245). The HiPurA® 96 Plasmid DNA Purification Kit works optimum when the same vector/host strain combination is used for all samples in a block prepared together. Growth of cultures varies depending on plasmid copy number, bacterial host strain, inoculation, antibiotic and type of culture medium. This may give rise to varied DNA concentrations in final samples. While using the HiPurA® 96 Plasmid DNA Purification Kit for the first time to prepare DNA for sequencing, it is recommended to prepare sequencing reactions using a range of different amounts of plasmid DNA to optimize quantity of template DNA for particular sequencing chemistry. This kit helps to recover upto 20 μg of high- or low-copy plasmid DNA from upto 2.0 ml of *E.coli* culture per isolation procedure.

Figure: Plasmid DNA obtained from 2.0 ml of recombinant *E.coli* culture using HiPurA® 96 Plasmid Purification Kit.
5μl of plasmid DNA sample was loaded on 0.8% agarose gel.



Samples: 1-28



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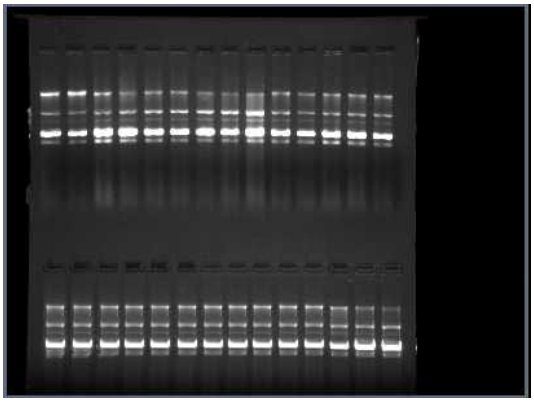

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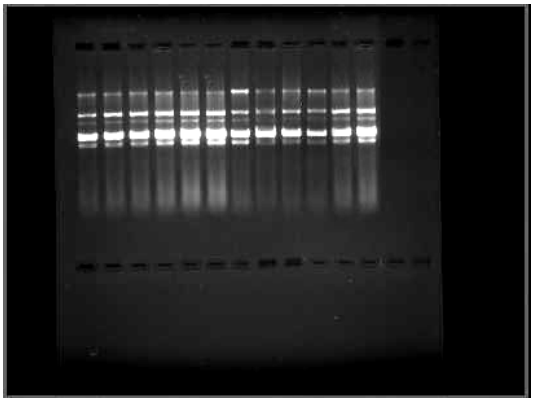

Samples: 85-96

HiPurA® 96-well DNA Plate (DBPL-96-01)

HiPurA® 96-well DNA Plate is based on the advanced silica binding principle presented in a centrifugation and vacuum format. The system efficiently couples the reversible nucleic acid-binding properties of the advanced silica membrane to yield high quantity of DNA. It facilitates the binding, washing and elution steps thus enabling multiple samples to be processed simultaneously. It eliminates the need for alcohol precipitation, expensive resins, and harmful organic compounds such as phenol and chloroform, otherwise employed in traditional DNA isolation techniques. DNA binds specifically to the advanced silica-gel membrane while

contaminants pass through. PCR inhibitors such as divalent cations and proteins are completely removed during wash steps, leaving pure nucleic acids to be eluted in the buffer provided with the kit. Plasmid DNA upto 20-30 kb in length can be purified for further downstream applications.

Elution

The yield of plasmid DNA depends on the copy number of the plasmid and the number of cells in the sample. A single elution with 150 µl of Elution Buffer (ET) will provide sufficient DNA to carry out multiple amplification reaction. Elution with volume less than 150 µl will increase the final DNA concentration, but will reduce the overall DNA yield.

Concentration, yield, and purity of DNA

Spectrophotometric analysis and agarose gel electrophoresis will reveal the concentration and the purity of the DNA. Use Elution Buffer (ET) to dilute samples and to calibrate the spectrophotometer, measure the absorbance at 260 nm, 280 nm and 320 nm using a quartz microcuvette. Absorbance readings at 260 nm should fall between 0.1 and 1.0. The 320 nm absorbance is used to correct for background absorbance. An absorbance of 1.0 at 260 nm corresponds to approximately 50 µg/ml of DNA. The A_{260}- A_{320} /A_{280}-A_{320} ratio should be 1.6–1.9. Purity is determined by calculating the ratio of absorbance at 260 nm to absorbance at 280 nm. DNA purified by HiPurA® 96 Plasmid DNA Purification Kit is free of protein and other contaminants that can inhibit enzymatic reactions or any downstream applications.

Concentration of DNA sample (µg/ml) = 50 x A_{260} x dilution factor.

Materials needed but not provided

- Tabletop Centrifuge with swinging bucket rotor and plate carriers, capable of atleast attaining 2,250 X g (≈3,700 rpm)
- Vacuum Manifold for 96 well plate
- Vacuum source regulator (pump) (capable of minimum -30 inches Hg pressure)
- Vacuum regulator
- Multi-channel pipette with tips
- Ethanol (96-100%)
- Oven preheated at 70°C

Storage

Store the HiPurA® 96 Plasmid DNA Purification Kit between 15-25°C except certain components as specified on each labels. Under recommended condition kit is stable for 1 year.

General Preparation Instructions

1. Add 5 µl RNase A (20 mg/ml) per 1ml of Resuspension Solution (HP1). After the addition of RNase A to HP1 it is stable for 6 months on storage at 2-8°C.

2. **Thoroughly mix reagents**
   Examine the solutions for any kind of precipitation. If any solution (except RNase A) forms a precipitate, warm at 55-65°C until the precipitate dissolves completely, allow it to cool to room temperature before use.

3. **Dilute Wash Solution Concentrate (HPE) (DS0024) as follows:**

Number of Preps	Wash Solution Concentrate (HPE)	Ethanol (96-100%)
1X96	40 ml	120 ml

User should carry out dilution of Wash Solution Concentrate (HPE) in a separate container, as it is not provided with this kit.

RNase A enzyme treatment

RNase A is a type of RNase that is commonly used in research for removal of RNA. RNase A (e.g., bovine pancreatic ribonuclease A) is one of the sturdiest enzymes in common laboratory usage. It cleaves 3' end of unpaired C and U residues.

Unit Definition for RNase A

One unit of the enzyme causes an increase in absorbance of 1.0 at 260nm when yeast RNA is hydrolyzed at 37°C and pH 5.0. Fifty units are approximately equivalent to 1 Kunitz unit. It is completely free of DNases and proteases. The specific activity is 90 U/mg. The product as supplied is stable at room temperature (15-25°C).

Centrifugation

All centrifugation steps are carried out in conventional laboratory centrifuge e.g. Beckman CS-6KR, Heraeus Varifuge 3.0R, or Sigma 6k10 with fixed angle rotor. The 96-well accessories provided with the kit are compatible with almost all laboratory centrifuges and rotors. All centrifugation steps are performed at room temperature and are given in g, the correct rpm can be calculated using the formula:

$$RPM = \sqrt{RCF/1.118 \times 10^{-5} r}$$

where *RCF* = required gravitational acceleration (relative centrifugal force in units of g); *r* = radius of the rotor in cm; and *RPM* = the number of revolutions per minute required to achieve the necessary *g*-force.

General Note:

1. **Vacuum protocol as well as centrifugation protocol can be performed for Plasmid DNA isolation. For a vacuum protocol, negative pressure of 30 mm Hg is required to achieve optimum results.**
2. **The HiPurA® Silicon Pad for sealing (PR11) used in the protocol should not be discarded. It can be reused after wiping with ethanol and washing appropriately.**

Specimen Collection and Handling

For Plasmid

Collect overnight culture from sterile flask with the help of micropipette. Store the remaining culture at 2-8°C for short term use.

Types of Specimen

Samples: Bacterial cultures

Procedure

Vacuum based protocol:

1. Inoculate each well of HiPurA® 96-well Block (2.2 ml) (LWB-96) with 1.3 ml growth medium containing appropriate selective antibiotic with a single bacterial colony. Carefully apply a sheet of Breathable Film (PR18) over the HiPurA® 96-well Block. Incubate the culture for 16-18 hours at 37°C at 250- 300 rpm.

NOTE: If required, wipe droplets if any from the top of the grow block before applying the Breathable Film (PR18).

2. Harvest the bacterial cells in the HiPurA® 96-well Block (2.2 ml) (LWB-96) by centrifugation at 5,000 rpm for 10 minutes in a centrifuge with a rotors and carriers for 96-well microplates. The block should be covered with breathable tape during centrifugation. Remove the medium by gently inverting the block onto a waste container.

   NOTE: Tap the inverted block gently onto an absorbent paper towel stack to drain out the remaining droplets of liquid. Ensure that the pellet is firm and does not get disturbed during the tapping.

3. **Resuspend Cells**
   Resuspend each bacterial pellet in 300 µl of Resuspension Solution (HP1) (DS0020) using multi- channel pipette for easy solution delivery. Seal the HiPurA® 96-well Block (2.2 ml) (LWB-96) using the provided HiPurA® Silicon Pad for sealing (PR11). Mix well by gentle vortexing till no cell clumps are visible. Remove the silicon pad from the top of the HiPurA® 96-well Block (2.2 ml) (LWB-96).

   NOTE: It is very important that homogenous suspension is obtained, as incomplete resuspension will result in poor recovery. Ensure that prior to use the appropriate amount of RNase A is added to HP1.

4. **Lyse Cells**
   Add 300 µl of Lysis Solution (HP2) (DS0021) to each well of the HiPurA® 96-well Block (2.2 ml) (LWB-96) to lyse the cells. Seal the HiPurA® 96-well Block (2.2 ml) (LWB-96) using the HiPurA® Silicon Pad provided with the kit (PR11). Mix thoroughly by gently inverting the 96-well Block (2.2 ml) 6-8 times.

   NOTE: It is important to mix gently when inverting the HiPurA® 96-well Block (2.2 ml) (LWB-96). Vigorous agitation may result in the shearing of genomic DNA, which may contaminate the plasmid DNA. Do not allow this lysis reaction to exceed more than 5 minutes. The bacterial suspension will become viscous after lysis. To avoid cross contamination between wells, centrifuge briefly at 5,000 rpm before proceeding with the next step. Allow the centrifuge to reach 5,000 rpm then stop. (Pulse spin)

5. **Neutralize**
   Add 450 µl of Neutralization Solution (HN3) (DS0022) to each well and reseal the HiPurA® 96-well Block (2.2 ml) (LWB-96) using the provided HiPurA® Silicon Pad for sealing (PR11). Mix gently and thoroughly by inverting the HiPurA® 96-well Block (2.2 ml) (LWB-96) 6-8 times.

   NOTE: The solution should become cloudy and the precipitation should be homogeneous.

6. Centrifuge the HiPurA® 96-well Block (2.2 ml) (LWB-96) at approximately 5,000 rpm for 10 minutes to obtain a compact white pellet.

   NOTE: A compact white pellet will form.

7. **Load lysate in HiPurA® 96-well DNA Plate**
   Connect the vacuum manifold to the vacuum source. Remove the manifold top and place a HiPurA® 96-well Block (1.6 ml) (LWB-16-96) into the manifold base to collect the flow-through liquid. Replace the manifold top and place the HiPurA® 96-well DNA Plate (DBPL-96-01) onto the manifold top.

8. Carefully remove the clear supernatant from HiPurA® 96-well Block (2.2 ml) (LWB-96) and transfer it to the corresponding wells in HiPurA® 96-well DNA Plate (DBPL-96-01). Place the HiPurA® Silicon Pad (PR11) to seal the plate.

   NOTE: Any Vacuum Manifold, which can accommodate 96-well plates, such as the Vacuum Manifold for 96 well plate can be used with the HiPurA® 96-well DNA Plate (DBPL-96-01). A negative pressure of -30 inches Hg is required. We recommend the use of a vacuum regulator to adjust the negative pressure. -30 inches Hg is equivalent to approximately 1000 mbar and 15 psi.

9. Turn on the vacuum source and adjust it to -30 inches Hg. Continue to draw vacuum through the plates until no liquid remains in any of the wells of the HiPurA® 96-well DNA Plate

10. Turn off the vacuum source and release the vacuum from inside of the vacuum manifold using the vacuum regulator. Remove the HiPurA® 96-well DNA Plate from the manifold and temporarily set it aside on a piece of absorbant toweling or plastic wrap.

    NOTE: Discard the flow-through liquid from HiPurA® 96-well Block (1.6 ml) (LWB-16-96) and reuse it (place it back into the manifold base).

11. **First wash**
    Place the HiPurA® 96-well DNA Plate onto the manifold top. Add 500 µl of Wash Solution (HPB) (DS0032) to each well of HiPurA® 96-well DNA Plate and place the HiPurA® Silicon Pad to seal (PR11) the plate. Turn on the vacuum source and adjust it to -30 inches Hg. Continue to draw vacuum until no Wash Solution is present in any of the wells. All the flow-through will be collected in the bottom HiPurA® 96-well Block (1.6 ml). Discard the flow-through liquid and reuse the block**.**

12. **Second wash**
    (Prepare Wash Solution as indicated in General Preparation Instructions)
    Place the HiPurA® 96-well DNA Plate (DBPL-96-01) onto the manifold top. Add 750 µl of diluted Wash Solution (HPE) (DS0024) to each well of HiPurA® 96-well DNA Plate. Apply vacuum until no Wash Solution is present in any of the wells. Repeat the wash one more time.

13. Discard the flow-through liquid and apply vacuum at a maximum of –30 inches Hg for 10 minutes to remove the traces of ethanol present in the Wash Solution.

14. Turn off the vacuum source. Remove the HiPurA® 96-well DNA Plate (DBPL-96-01) from the vacuum manifold and vigorously tap the plate approximately 6-8 times on several layers of absorbent toweling. Be careful not to damage the drip directors on the underside of the plate.

    NOTE: Lint-free absorbent toweling is recommended to avoid the release of tiny fibres, which could contaminate the plasmid and interfere with subsequent downstream applications.

15. Incubate HiPurA® 96-well DNA Plate open without the HiPurA® Silicon Pad for 15 minutes at 70°C in a vacuum oven to evaporate residual ethanol.

    NOTE: If vacuum oven is not available then connect the vacuum manifold to the oven pre-heated at 70°C through the port provided on the oven.

16. **DNA Elution**

Remove the HiPurA® 96-well Block (1.6 ml) (LWB-16-96) which was used as collection tray from the vacuum manifold base and replace it with a new HiPurA® 96-well V-plate for elution. Reassemble the manifold top and place the HiPurA® 96-well DNA Plate (DBPL-96-01) on the HiPurA® 96-well V-plate. Add 150 µl of Elution Buffer (ET) (DS0040) to the corresponding wells of HiPurA® 96-well DNA Plate (DBPL-96-01) and incubate at room temperature for 5 minutes. Turn on the vacuum source and allow the vacuum (-30 Hg pressure) to continue for 10 minutes.

17. Turn off the vacuum source and use the vacuum regulator to gradually release the vacuum from the manifold. Disassemble the manifold and remove the HiPurA® 96-well V-plate, which contains eluted plasmid DNA samples.

Centrifugation based protocol:

1. Inoculate each well of HiPurA® 96-well Block (2.2 ml) (LWB-96) with 1.3 ml growth medium containing appropriate selective antibiotic with a single bacterial colony. Apply carefully a sheet of Breathable Film (PR18) over the HiPurA® 96-well Block. Incubate the culture for 16-18 hours at 37°C at 250- 300 rpm.

   NOTE: If required, wipe droplets if any from the top of the grow block before applying the Breathable Film (PR18).

2. Harvest the bacterial cells in the HiPurA® 96-well Block (2.2 ml) (LWB-96) by centrifugation at 5,000 rpm for 10 minutes, in a centrifuge with a rotor and carriers for 96-well microplates. The block should be covered with breathable tape during centrifugation. Remove the medium by gently inverting the block onto a waste container.

   NOTE: Tap the inverted block gently onto an absorbent paper towel stack to drain out the remaining droplets of liquid. Ensure that the pellet is firm and does not get disturbed during the tapping.

3. **Resuspend Cells**
   Resuspend each bacterial pellet in 300 µl of Resuspension Solution (HP1) (DS0020) using multi- channel pipette for easy solution delivery. Seal the HiPurA® 96-well Block (2.2 ml) (LWB-96) using the provided HiPurA® Silicon Pad for sealing (PR11). Mix well by gentle vortexing till no cell clumps are visible. Remove the silicon pad from the top of the HiPurA® 96-well Block (2.2 ml) (LWB-96)

   NOTE: It is very important that homogenous suspension is obtained as incomplete resuspension results in poor recovery. Ensure that prior to use the appropriate amount of RNase A is added to HP1.

4. **Lyse Cells**
   Add 300 µl of Lysis Solution (HP2) (DS0021) to each well of the HiPurA® 96-well Block (2.2 ml) (LWB-96) to lyse the cells. Seal the HiPurA® 96-well Block (2.2 ml) (LWB-96) using the provided HiPurA® Silicon Pad for sealing (PR11). Mix thoroughly by gently inverting the 96-well Block (2.2 ml) 6-8 times.

   NOTE: It is important to mix gently when inverting the HiPurA® 96-well Block (2.2 ml) (LWB-96). Vigorous agitation may result in the shearing of genomic DNA, which may contaminate the plasmid DNA. Do not allow this lysis reaction to exceed for more than 5 minutes.

5. The bacterial suspension will become viscous after lysis. To avoid cross contamination between wells, centrifuge briefly at 5,000 rpm before proceeding with the next step. Allow the centrifuge to reach 5,000 rpm then stop. (Pulse spin)

6. **Neutralize**
   Add 450 µl of Neutralization Solution (HN3) (DS0022) to each well and reseal the HiPurA® 96-well Block (2.2 ml) (LWB-96) using the provided HiPurA® Silicon Pad for sealing (PR11). Mix gently and thoroughly by inverting the HiPurA® 96-well Block (2.2 ml) (LWB-96) six to eight times. Incubate the plate at room temperature for 10 minutes. Remove the HiPurA® Silicon Pad (PR11).

   NOTE: The solution should become cloudy and the precipitation should be homogeneous.

7. Centrifuge the HiPurA® 96-well Block (2.2 ml) (LWB-96) at approximately 5,000 rpm for 10 minutes to obtain a compact white pellet.

   NOTE: A compact white pellet will form.

8. **Load lysate in HiPurA® 96-well DNA Plate**
   Place the HiPurA® 96-well DNA Plate (DBPL-96-01) onto the HiPurA® 96-well Block (1.6 ml) (LWB-16-96). Carefully remove the supernatant from HiPurA® 96-well Block (2.2 ml) (LWB-96) and transfer it to the corresponding wells in HiPurA® 96-well DNA Plate (DBPL-96-01). Place the HiPurA® Silicon Pad for sealing (PR11) over it. Centrifuge the HiPurA® 96-well DNA Plate at 5,000 rpm for 5 minutes.

9. Discard the flow-through liquid from HiPurA® 96-well Block (1.6 ml) (LWB-16-96) and reuse it.

10. **First wash**
    Place the HiPurA® 96-well DNA Plate (DBPL-96-01) onto the HiPurA® 96-well Block. Add 500 µl of Wash Solution (HPB) (DS0032) to each well of HiPurA® 96-well DNA Plate (DBPL-96-01) and place the HiPurA® Silicon Pad for sealing (PR11) over it. Centrifuge at 5,000 rpm for 5 minutes. Discard the flow-through liquid and reuse the block.

11. **Second wash**
    (Prepare Wash Solution as indicated in General Preparation Instructions)
    Place the HiPurA® 96-well DNA Plate (DBPL-96-01) onto the HiPurA® 96-well Block. Add 750 µl of diluted Wash Solution (HPE) (DS0032) to each well of HiPurA® 96-well DNA Plate (DBPL-96-01). Centrifuge at approximately 5,000 rpm for 5 minutes. Repeat this wash step one more time.

12. Discard the flow-through liquid and centrifuge for 10 minutes at 5,000 rpm to remove the traces of ethanol present in the Wash Solution.

13. Incubate HiPurA® 96-well DNA Plate (DBPL-96-01) open for 15 minutes at 70°C in an incubator to evaporate residual ethanol.

14. **DNA Elution**
    Remove the HiPurA® 96-well Block (1.6 ml) (LWB-16-96), which was used as collection tray, and replace it with a new HiPurA® 96-well 1.6 ml block for elution. Place the HiPurA® 96-well DNA Plate (DBPL-96-01) on to it. Add 150 µl of Elution Buffer (ET) (DS0040) to the corresponding wells of HiPurA® 96-well DNA Plate (DBPL-96-01) and incubate at room temperature for 5 minutes. Centrifuge at approximately 5,000 rpm for 5 minutes.

15. Remove the HiPurA® 96-well Block (1.6 ml) (LWB-16-96), which contains eluted plasmid DNA samples.

Storage of the eluate with purified DNA: Transfer the eluted plasmid DNA to new 2 ml tubes. The eluate contains pure plasmid DNA. For short- term storage (24-48 hrs) of the DNA, 2-8°C is

recommended. For long-term storage, -20°C or lower temperature (-80°C) is recommended. Avoid repeated freezing and thawing of the sample as this may cause denaturing of DNA. The Elution Buffer helps to stabilize the DNA at these temperatures.

Warning and Precautions

Not for Medicinal Use. Read the procedure carefully before beginning the protocol. Wear protective gloves/protective clothing/eye protection/face protection. Follow good laboratory practices while handling samples. Standard precautions should be followed as per established guidelines. Safety guidelines may be referred in safety data sheets of the product.

Limitations

1. The yield of DNA depends upon the type and the volume of starting material used.

Performance and Evaluation

Each lot of HiMedia's HiPurA® 96 Plasmid DNA Purification Kit is tested against predetermined specifications to ensure consistent product quality.

Quality Control

Type of Sample	DNA Yield	DNA Purity
DH5α	upto 20 µg/ well	1.6-1.9

Reference

1. Sambrook, J., Fritsch, E.F., and Maniatis, T., *Molecular Cloning: A laboratory manual, Cold Spring Harbor Laboratory, second edition (1989).*

Troubleshooting Guide

Sr. No.	Problem	Cause	Solution
1.	Poor downstream enzymatic application	High salt concentration in the eluate	Ensure that the Wash Solution (HPE) wash step is carried out properly. Perform a second Wash Solution (HPE) wash step if necessary.
2.	Little/No DNA in eluate	Plasmid did not grow in culture	Ensure optimum growth conditions for the host strain with the plasmid. Check the age of clones and try again with freshly prepared single colonies or a new clone library.
		Incorrect preparation of lysates	Check the storage conditions and shelf-life of the buffers. Ensure addition of appropriate amount of buffers to the samples.
		Lysis Solution (HP2) precipitated	Redissolve the precipitate formed in Lysis Solution (HP2) by warming to 37°C (if required 55°C), before use.

		Incomplete Resuspension of cell	Completely resuspend cell pellet in Resuspension Solution (HP1). Do not add Lysis Solution (HP2) until cell pellet is resuspended completely.
3.	Poor quality DNA in eluate	RNA present as contamination in DNA eluate	Ensure that RNase A is added to Resuspension Solution (HP1). RNase A digestion should be sufficient. Add RNase A freshly to Resuspension Solution (HP1) before use. Reduce the culture volume if necessary to ensure complete RNase digestion.
4.	Genomic DNA contamination in plasmid eluate	Lysis Solution (HP2) added incorrectly	Handle lysate gently after addition of Lysis Solution (HP2) to avoid shearing of DNA. Culture volume may be reduced if lysate is too viscous for gentle mixing.
		Prolonged Lysis incubation time	The lysis incubation period in Step 4 should not exceed for more than 5 minutes.
		Culture has overgrown	Do not grow culture for longer than 20-24 hours.
5.	DNA sample oozes out from the well while loading on to the agarose gel	Residual ethanol present in the wash solution	After drying step, incubate the HiPurA® 96-well DNA Plate (DBPL-96-01) at 70°C for 15 minutes

Safety Information

The HiPurA® 96 Plasmid DNA Purification Kit is for laboratory use only, not for drug, household or other uses. Take appropriate laboratory safety measures and wear gloves when handling. Not compatible with disinfecting agents containing bleach. Please refer the Safety Data Sheet (SDS) for information regarding hazards and safe handling practices.

Disposal

User must ensure safe disposal by autoclaving and/or incineration of used or unusable preparations of this product. Follow established laboratory procedures in disposing of infectious materials and material that comes into contact with clinical sample must be decontaminated and disposed off in accordance with current laboratory techniques.

Technical assistance

At HiMedia, we pride ourselves on the quality and availability of our technical support. For any kind of technical assistance, mail to mb@himedialabs.com.

Storage temperature

Do not use if package is damaged

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02/2025

Disclaimer :

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