



# Technical Data

## Casitose Agar w/ 1.5% Agar

M793

### Intended Use

Recommended as a general purpose culture medium.

### Composition\*\*

Ingredients	Gms / Litre
Tryptone #	5.000
HM infusion B from ##	150.000
Peptone	5.000
Yeast autolysate	1.500
Sodium phosphate	2.500
Sodium chloride	5.000
Agar	15.000
Final pH ( at 25°C)	7.8±0.2

\*\*Formula adjusted, standardized to suit performance parameters

# Casein Hydrolysate

## Beef Infusion from

### Directions

Suspend 35.5 grams in 1000 ml purified / distilled water containing 22 ml glycerol. Heat to boiling to dissolve the medium completely. Sterilize by autoclaving at 15 lbs pressure (121°C) for 15 minutes. Cool to 45-50°C. Mix well and pour in sterile Petri plates.

### Principle And Interpretation

Casitose Agar w/1.5% is the modification of medium recommended by APHA (6) and is used as a general purpose culture medium.

It has casitose, HM infusion B from, and peptone which serves as a rich source of nitrogen and carbon. Yeast autolysate provides necessary growth factors and vitamin supplement required for metabolism of wide number of bacteria. Sodium phosphate helps buffering of media whereas sodium chloride balances the osmotic equilibrium.

### Type of Specimen

Food and dairy samples; Water samples

### Specimen Collection and Handling

For food and dairy samples, follow appropriate techniques for sample collection and processing as per guidelines (1,5,7).

For water samples, follow appropriate techniques for sample collection, processing as per guidelines and local standards.(2)

After use, contaminated materials must be sterilized by autoclaving before discarding.

### Warning and Precautions

Read the label before opening the container. Wear protective gloves/protective clothing/eye protection/ face protection. Follow good microbiological lab practices while handling specimens and culture. Standard precautions as per established guidelines should be followed while handling specimens. Safety guidelines may be referred in individual safety data sheets.

### Limitation

- 1.This medium is general purpose medium and may not support the growth of fastidious organisms.
2. Due to nutritional variations, some strains may show poor growth.

### Performance and Evaluation

Performance of the medium is expected when used as per the direction on the label within the expiry period when stored at recommended temperature.

## Quality Control

### Appearance

Yellow coloured homogeneous free flowing powder

### Gelling

Firm, comparable with 1.5% Agar gel

### Colour and Clarity of prepared medium

Light to medium amber coloured, clear to slightly opalescent gel forms in Petri plates.

### Reaction

Reaction of 3.55% w/v aqueous solution at 25°C. pH :  $7.8 \pm 0.2$

### pH

7.60-8.00

### Cultural Response

Cultural characteristics observed after an incubation at 35-37°C for 18-24 hours .

Organism	Inoculum (CFU)	Growth	Recovery
<i>Enterococcus faecalis</i> ATCC 29212 (00087*)	50-100	luxuriant	$\geq 70\%$
<i>Staphylococcus aureus</i> subsp. <i>aureus</i> ATCC 25923 (00034*)	50-100	luxuriant	$\geq 70\%$

Key : \*Corresponding WDCM numbers.

## Storage and Shelf Life

Store between 10-30°C in a tightly closed container and the prepared medium at 20-30°C. Use before expiry date on the label. On opening, product should be properly stored dry, after tightly capping the bottle in order to prevent lump formation due to the hygroscopic nature of the product. Improper storage of the product may lead to lump formation. Store in dry ventilated area protected from extremes of temperature and sources of ignition. Seal the container tightly after use.

Product performance is best if used within stated expiry period.

## Disposal

User must ensure safe disposal by autoclaving and/or incineration of used or unusable preparations of this product. Follow established laboratory procedures in disposing of infectious materials and material that comes into contact with sample must be decontaminated and disposed of in accordance with current laboratory techniques (3,4).

## Reference

1. American Public Health Association, Standard Methods for the Examination of Dairy Products, 1978, 14th Ed., Washington D.C.
2. Baird R.B., Eaton A.D., and Rice E.W., (Eds.), 2015, Standard Methods for the Examination of Water and Wastewater, 23rd ed., APHA, Washington, D.C.
3. Isenberg, H.D. Clinical Microbiology Procedures Handbook 2<sup>nd</sup> Edition.
4. Jorgensen, J.H., Pfaller, M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
5. Salfinger Y., and Tortorello M.L., 2015, Compendium of Methods for the Microbiological Examination of Foods, 5th Ed., American Public Health Association, Washington, D.C.
6. Vanderzant C and Splittstoesser D (Eds) 1992. Compendium of Methods for the Microbiological Examination of Foods, 3rd ed, APHA, Washington, DC.
7. Wehr H. M. and Frank J. H., 2004, Standard Methods for the Microbiological Examination of Dairy Products, 17th Ed., APHA Inc., Washington, D.C.

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