



## Lysine Decarboxylase Broth

M376

### Intended Use:

Recommended for differentiating *Salmonella arizonae* from the Bethesda Ballerup group of *Enterobacteriaceae*.

### Composition\*\*

Ingredients	g / L
Peptone	5.000
Yeast extract	3.000
Dextrose (Glucose)	1.000
L-Lysine hydrochloride	5.000
Bromocresol purple	0.020
Final pH ( at 25°C)	6.8±0.2

\*\*Formula adjusted, standardized to suit performance parameters

### Directions

Suspend 14.02 grams in 1000 ml purified / distilled water. Heat, if necessary to dissolve the medium completely. Dispense 5 ml amount into screw-capped test tubes. Sterilize by autoclaving at 15 lbs pressure (121°C) for 15 minutes. Cool the tubed medium in an upright position and overlay with 2-3 ml of sterile mineral oil.

### Principle And Interpretation

Decarboxylase media were first described by Moeller (1,2,3) for detecting lysine and ornithine decarboxylase and arginine dihydrolase. Falkow developed a lysine decarboxylase medium for the identification and differentiation of *Salmonella* and *Shigella* (4). Lysine Decarboxylase Broth is especially suited to study the decarboxylase reactions for members of *Enterobacteriaceae* other than *Klebsiella* and *Enterobacter*. Lysine Decarboxylase Broth is also recommended by APHA (5,6) and other standard methods (5,7).

During the initial stages of incubation, following inoculation, fermentation of dextrose by the organisms leads to acid production, which causes a subsequent colour change of the bromocresol purple indicator to yellow. The acidic condition thus generated stimulates decarboxylase activity, which leads to decarboxylation of lysine to cadaverine. The alkaline conditions generated due to cadaverine production cause the bromocresol purple indicator (changed to yellow) to revert to purple colour. If the organisms do not produce decarboxylase enzyme, the colour of the medium remains yellow. Dextrose non-utilizers will not show any change in the medium colour. Use light inocula and do not read the tests after 24 hours incubation, as some organisms require longer incubation time of upto 4 days.

### Type of specimen

Clinical samples, Food samples; Water samples.

### Specimen Collection and Handling:

For clinical samples follow appropriate techniques for handling specimens as per established guidelines (4,8).

For food samples, follow appropriate techniques for sample collection and processing as per guidelines (7). For water samples, follow appropriate techniques for sample collection, processing as per guidelines and local standards. (6) After use, contaminated materials must be sterilized by autoclaving before discarding.

### Warning and Precautions :

In Vitro diagnostic device. For professional use only. Read the label before opening the container. Wear protective gloves/protective clothing/eye protection/ face protection. Follow good microbiological lab practices while handling specimens and culture. Standard precautions as per established guidelines should be followed while handling specimens. Safety guidelines may be referred in individual safety data sheets.

## Limitations:

1. Use light inoculum and do not read the tests after 24 hours incubation, as some organisms require longer incubation time of upto 4 days.
2. Other biochemical and serological tests must be performed in conjunction for confirmation.

## Performance and Evaluation

Performance of the medium is expected when used as per the direction on the label within the expiry period when stored at recommended temperature.

## Quality Control

### Appearance

Light yellow to greenish yellow homogeneous free flowing powder

### Colour and Clarity of prepared medium

Purple coloured clear solution without any precipitate

### Reaction

Reaction of 1.4% w/v aqueous solution at 25°C. pH : 6.8±0.2

### pH

6.60-7.00

### Cultural Response

Cultural characteristics observed after an incubation at 35-37°C for 18-24 hours. (Inoculated tubes are overlayed with sterile mineral oil).

Organism	Lysine decarboxylation
<i>Citrobacter freundii</i> ATCC 8090	variable reaction
<i>Escherichia coli</i> ATCC 25922 (00013*)	variable reaction
# <i>Klebsiella aerogenes</i> ATCC 13048 (00175*)	positive reaction, purple colour
<i>Klebsiella pneumoniae</i> ATCC 13883 (00097*)	positive reaction, purple colour
<i>Proteus mirabilis</i> ATCC 25933	negative reaction, yellow colour
\$ <i>Proteus hauseri</i> ATCC 13315	negative reaction, yellow colour
<i>Salmonella Arizonae</i> ATCC 13314	Positive reaction, purple colour
<i>Salmonella</i> Paratyphi A ATCC 9150	negative reaction, yellow colour
<i>Salmonella</i> Typhi ATCC 6539	positive reaction, purple colour
<i>Serratia marcescens</i> ATCC 8100	positive reaction, purple colour
<i>Shigella dysenteriae</i> ATCC 13313	negative reaction, yellow colour

Key : (\*) Corresponding WDCM numbers.

(#) Formerly known as *Enterobacter aerogenes*

\$ Formerly known as *Proteus vulgaris*

## Storage and Shelf Life

Store between 10-30°C in a tightly closed container and the prepared medium at 15-30°C. Use before expiry date on the label. On opening, product should be properly stored dry, after tightly capping the bottle in order to prevent lump formation due to the hygroscopic nature of the product. Improper storage of the product may lead to lump formation. Store in dry ventilated area protected from extremes of temperature and sources of ignition. Seal the container tightly after use. Product performance is best if used within stated expiry period.

## Disposal

User must ensure safe disposal by autoclaving and/or incineration of used or unusable preparations of this product. Follow established laboratory procedures in disposing of infectious materials and material that comes into contact with sample must be decontaminated and disposed of in accordance with current laboratory techniques (4,8).

## References

1. Moeller V., 1954, Acta. Pathol. Microbiol. Scand., 34:102.
2. Moeller V., 1954, Acta. Pathol. Microbiol. Scand., 34:259.
3. Moeller V., 1955, Acta. Pathol. Microbiol. Scand., 36:158. 4..Falkow, 1958, Am. J. Clin. Pathol., 29:598.Isenberg, H.D. Clinical Microbiology Procedures Handbook 2<sup>nd</sup> Edition.
4. Jorgensen, J.H., Pfaller, M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
5. FDA Bacteriological Analytical Manual, 2017, AOAC, Washington, DC.
6. Lipps WC, Braun-Howland EB, Baxter TE,eds. Standard methods for the Examination of Water and Wastewater, 24th ed. Washington DC:APHA Press; 2023.
7. Salfinger Y., and Tortorello M.L., 2015, Compendium of Methods for the Microbiological Examination of Foods,5th Ed., American Public Health Association, Washington, D.C.
8. Isenberg, H.D. Clinical Microbiology Procedures Handbook 2nd Edition.

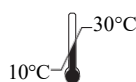
Revision : 04/ 2024



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