



## Wild Yeast Medium

### Intended Use:

Recommended for detection of wild yeast.

### Composition\*\*

Ingredients	Gms / Litre
Peptone	5.000
Yeast extract	3.000
Malt extract	3.000
Dextrose (Glucose)	10.000
Basic fuchsin	0.470
Sodium sulphite	2.920
Dextrin	0.110
Crystal violet	0.001
Agar	20.000
Final pH ( at 25°C)	6.9±0.2

\*\*Formula adjusted, standardized to suit performance parameters

### Directions

Suspend 44.50 grams in 1000 ml purified / distilled water. Boil with constant stirring for 15 minutes. DO NOT AUTOCLAVE. Cool to 45-50°C mix well and pour into sterile Petri plates. Efficacy of the plates can be improved by incubating them to 30°C for 18 hours before use.

### Principle And Interpretation

Wild Yeast Medium is used for the detection of *Saccharomyces* wild yeast. Malt extract, Peptone and yeast extract provide carbon, nitrogen compounds, long chain amino acids, vitamins and other necessary nutrients to support the growth of yeasts. Dextrose is the suitable carbohydrate for the growth of yeasts. Sodium sulphite and basic fuchsin inhibit the gram-positive microorganisms.

The prepared plates darken during incubation. Wild yeasts form pink colonies which may be smooth, mucoid or wrinkled. Brewing yeasts forms a thin haze of micro colonies which blend with the colour of the medium.

### Type of specimen

Brewing yeasts

### Specimen Collection and Handling

For Brewing yeast samples, follow appropriate techniques for sample collection, processing as per guidelines and local standards (3).

After use, contaminated materials must be sterilized by autoclaving before discarding.

### Warning and Precautions

Read the label before opening the container. Wear protective gloves/protective clothing/eye protection/ face protection. Follow good microbiological lab practices while handling specimens and culture. Standard precautions as per established guidelines should be followed while handling specimens. Safety guidelines may be referred in individual safety data sheets.

### Limitations

1. This medium is general purpose medium and may not support the growth of fastidious organisms.

## Performance and Evaluation

Performance of the medium is expected when used as per the direction on the label within the expiry period when stored at recommended temperature.

## Quality Control

### Appearance

Pinkish purple to purple homogeneous free flowing powder

### Gelling

Firm, comparable with 2.0% agar gel.

### Colour and Clarity

Light pink coloured clear to slightly opalescent gel forms in Petri plates.

### Reaction

Reaction of 4.45% aqueous solution at 25°C. pH : 6.9±0.2

### Cultural Response

Cultural response observed at 30°C for upto 4 days.(colour of plates darkens during incubation).

Organism	Inoculum (CFU)	Growth	Recovery	Colour of Colony
<i>Candida albicans</i> ATCC 10231 (00054*)	50-100	luxuriant	≥50%	white to light pink raised colonies
<i>Candida krusei</i> ATCC 24408	50-100	luxuriant	≥50%	pink, rough, flat colonies
<i>Saccharomyces cerevisiae</i> ATCC 9763 (00058*)	50-100	luxuriant	≥50%	pink colonies

Key : (\*) Corresponding WDCM numbers.

## Storage and Shelf Life

Store between 10-30°C in a tightly closed container and the prepared medium at 20-30°C. Use before expiry date on the label. On opening, product should be properly stored dry, after tightly capping the bottle in order to prevent lump formation due to the hygroscopic nature of the product. Improper storage of the product may lead to lump formation. Store in dry ventilated area protected from extremes of temperature and sources of ignition. Seal the container tightly after use. Product performance is best if used within stated expiry period.

## Disposal

User must ensure safe disposal by autoclaving and/or incineration of used or unusable preparations of this product. Follow established laboratory procedures in disposing of infectious materials and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (1,2).

## Reference

1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2<sup>nd</sup> Edition.
2. Jorgensen, J.H., Pfaller, M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
3. Pelczar M.J.Jr., Reid R.D., Chan E. C.S,1977, Microbiology, 4th ed, Tata „McGraw Hill Publishing company limited, New Delhi.

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### Disclaimer :

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