

Technical Data

HiCrome[™] Selective Salmonella Agar Base

M1842

Intended Use:

Recommended for the selective isolation of Salmonella species from food and clinical samples.

Composition**

Ingredients	g/L
HI powder #	12.000
Yeast hydrolysate	5.000
Tryptose	5.000
Sodium cholate	3.000
Sodium taurocholate	5.000
Sodium deoxycholate	1.000
Chromogenic mixture	8.000
Agar	15.000
Final pH (at 25°C)	7.3 ± 0.2

^{**}Formula adjusted, standardized to suit performance parameters

Directions

Suspend 54.00 gram in 1000 ml purified/ distilled water. Gently heat to boiling to dissolve the medium completely. **DO NOT AUTOCLAVE.** Cool to 45-50°C. Aseptically add the rehydrated contents of one vial of NC Selective Supplement (FD274). Mix well and pour into sterile Petri plates.

Principle And Interpretation

Salmonella species have been isolated from humans and almost all animals throughout the world. They cause many types of infections from mild, self-limiting gastroenteritis to life threatening typhoid fever. Salmonella Typhi and Salmonella Paratyphi A & B cause gastroenteritis, bacteremia and enteric fever, Salmonella Choleraesuis causes gastroenteritis and enteric fever, especially in children. Salmonella Typhimurium is the most frequently isolated serotype of Salmonella. Salmonella species are the major cause of food poisoning (1).

Various chromogenic media are available for the differentiation of *Salmonella* species. The original media formulated by Rambach (2) differentiates *Salmonella* based on propylene glycol utilization and presence of a chromogenic indicator. However HiCromeTM Selective Salmonella Agar Base uses chromogenic mixture for identification and differentiation of *Salmonella* species. Sodium cholate, Sodium taurocholate and Sodium deoxycholate in the medium helps to restrict the growth of other organisms. Besides the selective supplement added to the medium inhibits competing microorganisms.

HI powder, yeast hydrolysate and tryptose in the medium provides nitrogenous, carbonaceous compounds, long chain amino acids, vitamins and other essential growth nutrients. Due to the presence of chromogenic mix in the medium *Salmonella* are easily distinguishable and forms purple coloured colonies while some *Enterobacteriaceae* like *Klebsiella* and *Enterobacter* forms blue to dark blue coloured colonies.

Conventional method employes the H₂S production property for *Salmonella* detection which is also exhibited by other non *Salmonella* species such as *Citrobacter*, *Proteus* etc. Hence further biochemical confirmation is required for further identification. This medium is specially employed for food samples where the sample is initially enriched in *Salmonella* Selective Enrichment Broth (M1843) and then isolated on HiCromeTM Selective Salmonella Agar Base. *Salmonella* species give purple coloured colonies due to the enzyme specificity.

Type of specimen

Clinical samples- stool, urine, etc; Food samples

[#] Equivalent to Heart Infusion powder

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Specimen Collection and Handling

For clinical samples follow appropriate techniques for handling specimens as per established guidelines (3.4).

For food samples, follow appropriate techniques for sample collection and processing as per guidelines (5).

After use, contaminated materials must be sterilized by autoclaving before discarding.

Warning and Precautions

In Vitro diagnostic use. For professional use only. Read the label before opening the container. Wear protective gloves/protective clothing/eye protection/face protection. Follow good microbiological lab practices while handling specimens and culture. Standard precautions as per established guidelines should be followed while handling clinical specimens. Safety guidelines may be referred in individual safety data sheets.

Limitations

- 1. Being highly selective, some strains may show poor growth.
- 2. Most of the Salmonella strains shows purple colonies except few.
- 3. Final confirmation of suspected colonies must be carried out by serological and biochemical tests.

Performance and Evaluation

Performance of the medium is expected when used as per the direction on the label within the expiry period when stored at recommended temperature.

Quality Control

Appearance

Light yellow to beige homogeneous free flowing powder

Gelling

Firm, comparable with 1.5 % Agar gel.

Colour and Clarity of prepared medium

Whitish cream coloured, opaque gel forms in Petri plates

Reaction

Reaction of 5.4% w/v aqueous solution at 25°C. pH: 7.3±0.2

pН

7.10 - 7.50

Cultural Response

Cultural characteristics observed with added NC Selective Supplement (FD274), after an incubation at 35-37°C for 22-24 hours.

Inoculum (CFU)	Growth	Recovery	Colour of colony
>=104	inhibited	0%	
50 -100	good	40 -50 %	blue
50 -100	good-luxuriant	>=50 %	purple
50 -100	good-luxuriant	>=50 %	purple
>=104	inhibited	0 -0 %	
	(CFU) >=10 ⁴ 50 -100 50 -100	(CFU) >=10 ⁴ inhibited 50 -100 good 50 -100 good-luxuriant 50 -100 good-luxuriant	(CFU) >=10 ⁴ inhibited 0% 50 -100 good 40 -50 % 50 -100 good-luxuriant >=50 % 50 -100 good-luxuriant >=50 %

Key: (*) Corresponding WDCM numbers

Storage and Shelf Life

Store dehydrated medium in a tightly closed container and the prepared medium at 2-8°C. Use before expiry date on the label. On opening, product should be properly stored dry, after tightly capping the bottle in order to prevent lump formation due to the hygroscopic nature of the product. Improper storage of the product may lead to lump formation. Store in dry ventilated area protected from extremes of temperature and sources of ignition Seal the container tightly after use. Product performance is best if used within stated expiry period.

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Disposal

User must ensure safe disposal by autoclaving and/or incineration of used or unusable preparations of this product. Follow established laboratory procedures in disposing of infectious materials and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (3,4).

Reference

- 1. Murray P. R., Baron J. H., Pfaller M. A., Jorgensen J. H. and Yolken R. H., (Ed.), 2003, Manual of Clinical Microbiology, 8th Ed., American Society for Microbiology, Washington, D.C.
- 2. Rambach A., 1990, Appl. Environ. Microbiol., 56:301.
- 3. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition.
- 4.Jorgensen, J.H., Pfaller, M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
- 5. Salfinger Y., and Tortorello M.L., 2015, Compendium of Methods for the Microbiological Examination of Foods, 5th Ed., American Public Health Association, Washington, D.C.

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In vitro diagnostic medical device





Storage temperature



Do not use if package is damaged

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