



## Mueller Kauffman Tetrathionate Novobiocin Broth Base

M1496I

### Intended Use:

Recommended for improved enrichment and isolation of *Salmonellae*. The composition and performance criteria of this media are as per the specification laid down in ISO 6579-1:2017 and ISO 11133:2014 (E) /Amd. : 2020.

### ISO Specification - Muller-Kauffmann tetrathionate-novobiocin (MKTTn) broth

### M1496I - Mueller Kauffman Tetrathionate Novobiocin Broth Base

### Composition\*\*

Ingredients	g / L	Ingredients	g / L
Meat extract	4.300	HM extract#	4.300
Enzymatic digest of casein	8.600	Tryptone###	8.600
Ox bile for bacteriological use	4.780	Bile##	4.780
Sodium chloride (NaCl)	2.600	Sodium chloride	2.600
Calcium carbonate (CaCO <sub>3</sub> )	38.700	Calcium carbonate	38.700
Sodium thiosulphate, pentahydrate (Na <sub>2</sub> S <sub>2</sub> O <sub>3</sub> · 5H <sub>2</sub> O)	47.800	Sodium thiosulphate, pentahydrate	47.800
Brilliant green	0.0096	Brilliant green	0.0096
Final pH ( at 25°C)	8.0±0.2	Final pH ( at 25°C)	8.0±0.2

### Supplements to be added after autoclaving

Novobiocin sodium salt 0.040

**Iodine-iodide solution** 20.00ml  
Iodine 4.000  
Potassium iodide (KI) 5.000

### MKTT Supplement

### FD203

Novobiocin 0.040

**\$ Iodine-iodide solution** 20.000ml  
Iodine 4.000  
Potassium iodide (KI) 5.000

\*\*Formula adjusted, standardized to suit performance parameters

# Equivalent to Meat extract

## Equivalent to Ox bile

### Equivalent to Enzymatic digest of casein

\$ To be added but not provided (To be freshly prepared)

### Directions

Suspend 89.42 gram (equivalent weight of dehydrated medium per litre) in 1000 ml purified/ distilled water. Heat the medium just to boiling. DO NOT AUTOCLAVE. Cool to 45-50°C and just before use aseptically add rehydrated contents of 1 vial of MKTT Novobiocin Supplement (FD203) and 20 ml of iodine-iodide solution (20 gram iodine and 25 gram potassium iodide in 100 ml sterile distilled water). Mix well to disperse calcium carbonate uniformly before dispensing in sterile tubes.

*Note: Due to presence of calcium carbonate, the prepared media forms opalescent solution with white precipitate.*

### Principle And Interpretation

The examination of various types of food products for *Salmonella* requires methods different from those used in clinical laboratories. The need for such method is due to the generally low numbers of *Salmonellae* in foods and the frequently poor physiological state of these pathogens following exposure to stressful conditions during food processing or storage. Injured *Salmonella* are resuscitated in non-selective broth medium, which facilitates detection of sublethally injured *Salmonella*. The ideal pre-enrichment broth should provide for the repair of cell damage, dilute toxic or inhibitory substances and nutritive enough to favour growth of *Salmonella*.

Mueller (1) recommended Tetrathionate Broth as a selective medium for the isolation of *Salmonella*. Kauffman (2) modified the formula to include ox bile and brilliant green as selective agents to suppress bacteria such as *Proteus* species. The British Standard Specification specifies Brilliant Green Tetrathionate Broth for isolating *Salmonella* from meat, meat products, and from poultry and poultry products (3). ISO committee has also recommended this pre-enrichment medium for the detection of *Salmonella* species from food stuffs and other materials (4). Selectivity is conferred by tetrathionate (from the reaction of thiosulphate and iodine).

Using more than one selective broth increases the isolation of *Salmonella* from samples with multiple serotypes (1). This medium contains Tryptone and HM extract as sources of carbon, nitrogen, vitamins and minerals. Bile and added brilliant green are selective agents, which inhibit gram-positive and other gram-negative organisms. Calcium carbonate is the buffer. Sodium chloride maintains osmotic equilibrium. Sodium thiosulphate is a source of sulfur. The tetrathionate (S<sub>4</sub>O<sub>6</sub>) anions constitute the principle selective agent in these enrichment media. Organisms other than *Salmonellae*, such as *Morganella morganii* and some *Enterobacteriaceae* may grow in the medium. Therefore, confirmatory tests should be carried out on all presumptive *Salmonella* colonies that are recovered. Method (5).

### Type of specimen

Food samples including milk and milk products, in animal feed, in animal faeces, and in environmental samples from the primary production stage.

### Specimen Collection and Handling:

**Processsing : ISO 6579-1:2017 (4)**

**Pre-enrichment :** Samples (25 gram in 225 ml) are preenriched in Buffered Peptone Water (M1494I) and incubated at 34° C to 38°C for 18 h ± 2 hours.

**Selective enrichment:** 0.1 ml of pre- enriched sample is inoculated in 10 ml RVS Broth (M1448I) or MSRV Agar (M1428I) and incubated at 41.5 ± 1°C for 24 ± 3 hours and 1 ml of culture is inoculated in MKTTn broth (M1496I) and incubated at 37± 1°C for 24 ± 3 hours .

**Isolation :** The culture thus obtained is then plated on XLD Agar, Modified (M031I) and incubated at 37± 1°C for 24 ± 3 hours . Simultaneously plating on second isolation agar is carried out.

**Confirmation :** Biochemical and serological tests are performed for confirmation.

### Warning and Precautions :

Read the label before opening the container. Wear protective gloves/protective clothing/eye protection/ face protection.

Follow good microbiological lab practices while handling specimens and culture. Standard precautions as per established guidelines should be followed while handling specimens. Safety guidelines may be referred in individual safety data sheets.

### Limitations :

- 1.The complete medium is unstable and should be used immediately. After incubation, it is permissible to store the selective enrichment medium at 5°C for a maximum of 72 h.
- 2.Individual organisms differ in their growth requirement and may show variable growth patterns in the medium
- 3.Each lot of the medium has been tested for the organisms specified on the COA. It is recommended to users to validate the medium for any specific microorganism other than mentioned in the COA based on the user's unique requirement.
- 4.Confirmatory tests should be carried out on all presumptive *Salmonella* colonies that are recovered.

### Performance and Evaluation

Performance of the medium is expected when used as per the direction on the label within the expiry when stored at period recommended temperature.

### Quality Control

#### Appearance

Cream to greenish yellow homogeneous free flowing powder

#### Colour and Clarity of prepared medium

Light green coloured opalescent solution forms with heavy white precipitate

#### Reaction

Reaction of 8.93% w/v aqueous solution at 25°C. pH : 8.0±0.2

#### pH

7.80-8.20

#### Cultural Response

**Productivity:** Cultural response was observed after an incubation at 37 ±1°C for 24 ± 3 hours with added sterile 20ml iodine solution and MKTT Supplement (FD203). Further subculture is carried out on XLD Agar, Modified (M031I) and incubated at 37 ±1°C for 24 ± 3 hours.

Organism	Inoculum (CFU)	Recovery on XLD Agar (M031I)	Colour of colony on XLD Agar (M031I)
<i>Salmonella Enteritidis</i> ATCC 13076 (00030*)	50-100	>10 colonies	red colonies w/ black centre
+ <i>Escherichia coli</i> ATCC 8739 (00012*)	$\geq 10^4$		
+ <i>Pseudomonas aeruginosa</i> ATCC 27853 (00025*)	$\geq 10^4$		
<i>Salmonella Typhimurium</i> ATCC 14028 (00031*)	50-100	>10 colonies	red colonies w/ black centre
+ <i>Escherichia coli</i> ATCC 25922 (00013*)	$\geq 10^4$		
+ <i>Pseudomonas aeruginosa</i> ATCC 27853 (00025*)	$\geq 10^4$		

**Selectivity :** Cultural characteristics observed after an incubation at  $37 \pm 1^\circ\text{C}$  for  $24 \pm 3$  hours with added sterile 20ml iodine solution and MKTT Supplement (FD203). Further subculture is carried out on Tryptone Soya Agar (M290) and incubated at  $37 \pm 1^\circ\text{C}$  for  $24 \pm 3$  hours.

Organism	Inoculum (CFU)	Growth	Recovery on Tryptone Soya Agar
<i>Escherichia coli</i> ATCC 8739 (00012*)	$\geq 10^4$	partial inhibition	$\leq 100$ colonies
<i>Escherichia coli</i> ATCC 25922 (00013*)	$\geq 10^4$	partial inhibition	$\leq 100$ colonies
<i>Enterococcus faecalis</i> ATCC 29212 (00087*)	$\geq 10^4$	inhibition - partial inhibition	<10 colonies
<i>Enterococcus faecalis</i> ATCC 19433 (00009*)	$\geq 10^4$	inhibition - partial inhibition	<10 colonies

Key: (\*) - Corresponding WDCM Numbers

## Storage and Shelf Life

Store between  $10-30^\circ\text{C}$  in a tightly closed container and use freshly prepared medium. Use before expiry date on the label. On opening, product should be properly stored dry, after tightly capping the bottle in order to prevent lump formation due to the hygroscopic nature of the product. Improper storage of the product may lead to lump formation. Store in dry ventilated area protected from extremes of temperature and sources of ignition. Seal the container tightly after use. Product performance is best if used within stated expiry period.

## Disposal

User must ensure safe disposal by autoclaving and/or incineration of used or unusable preparations of this product. Follow established laboratory procedures in disposing of infectious materials and material that comes into contact with sample must be decontaminated and disposed of in accordance with current laboratory techniques (6,7).

## Reference

1. Mueller L., 1923, C. R. Soc. Biol., (Paris) 89:434. Harvey R. W. S. and Price T. S., 1976, J. Hyg. Camb., 77:333.
2. Kauffman F., 1935, Ztschr. F. Hyg., 117:26.
3. Public Health Laboratory Service, 1974, Monograph Series No. 8, Public Health Laboratory Service, London, England.
4. Microbiology of the food chain — Horizontal method for the detection, enumeration and serotyping of *Salmonella* — Detection of *Salmonella* spp. ISO 6579-1:2017
5. Microbiology of food, animal feeding stuffs and water- Preparation, production, storage and performance testing of culture media, EN ISO 11133:2014 (E) /Amd. :2020 .
6. Isenberg, H.D. Clinical Microbiology Procedures Handbook 2nd Edition.
7. Jorgensen, J.H., Pfaller, M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.

Revision : 06/2024

**Disclaimer :**

User must ensure suitability of the product(s) in their application prior to use. Products conform solely to the information contained in this and other related HiMedia™ publications. The information contained in this publication is based on our research and development work and is to the best of our knowledge true and accurate. HiMedia™ Laboratories Pvt Ltd reserves the right to make changes to specifications and information related to the products at any time. Products are not intended for human or animal or therapeutic use but for laboratory, diagnostic, research or further manufacturing use only, unless otherwise specified. Statements contained herein should not be considered as a warranty of any kind, expressed or implied, and no liability is accepted for infringement of any patents.