



Modified Rappaport Vassiliadis Medium

M1137

Intended Use:

Recommended for selective enrichment of Salmonellae from food and environmental specimens. **Composition****

Ingredients	Gms / 1110 ml
Soya peptone	5.000
Sodium chloride	8.000
Potassium dihydrogen phosphate	1.600
Magnesium chloride hexahydrate	40.000
Malachite green	0.040
Final pH (at 25°C)	5.2±0.2
Example adjusted standardized to quit nonfermance nonemate	*

**Formula adjusted, standardized to suit performance parameters

Directions

Suspend 30.07 grams (the equivalent weight of dehydrated medium per litre) in 1000 ml purified/distilled water. Heat gently if necessary to dissolve the medium completely. Dispense in tubes or flasks as desired. Sterilize by autoclaving at Δ 115°C for 15 minutes.

 Δ corresponds to 10 lbs pressure.

Principle And Interpretation

Modified Rappaport Vassiliadis Medium is a selective broth for the enrichment of *Salmonella* from foodstuffs, environment and clinical specimens. The original formulation described by Rappaport et al (3) with magnesium chloride hexahydrate was modified by Vassiliadis et al (4) by lowering the concentration of malachite green and raising the incubation temperature to 43°C. This medium is recommended as the selective enrichment medium for isolation *of Salmonella* from food and environmental specimens.

Soya peptone serves as a source of nitrogen and carbon compounds, long chain amino acids, vitamins and other essential nutrients. Phosphates buffer the medium. Magnesium chloride raises the osmotic pressure in the medium. Malachite green inhibits gram positive bacteria and selectively enriches growth of *Salmonella* species. Sodium chloride maintains the osmotic balance.

Type of specimen

Food samples and Environmental samples

Specimen Collection and Handling:

The test specimen is added to Buffered Peptone Water (M614) and incubated at 35° C for 16 - 20 hours. This pre-enriched peptone water culture is inoculated into Modified Rappaport Vassiliadis Medium and incubated at $42 \pm 1^{\circ}$ C for 24 - 48 hours and further subcultured on Brilliant Green Agar (M016). After use, contaminated materials must be sterilized by autoclaving before discarding.

Warning and Precautions :

Read the label before opening the container. Wear protective gloves/protective clothing/eye protection/face protection. Follow good microbiological lab practices while handling specimens and culture. Standard precautions as per established guidelines should be followed while handling specimens. Safety guidelines may be referred in individual safety data sheets.

Limitations

Further biochemical and serological tests must be carried out for further identification.

2. Individual organisms differ in their growth requirement and may show variable growth patterns on the medium.

3. Each lot of the medium has been tested for the organisms specified on the COA. It is recommended to users to validate the medium for any specific microorganism other than mentioned in the COA based on the user's unique requirement.

3HUIRUPDQFH DQG (YDOXDWLRQ

3HUIRUPDQFH RIWKH PHGLXPLVH[SHFWHG ZKHQ XHYSHLGUDSHSUHLUROW KZHKHGQLUVH) UHFRPPHQGHG WHPSHUDWXUH

Quality Control

Appearance

Light yellow to light blue homogeneous free flowing powder

Colour and Clarity of prepared medium

Blue coloured clear solution without any precipitate

Reaction

Reaction of 3.0% w/v aqueous solution at 25°C. pH : 5.2±0.2

pН

5.00-5.40

Cultural Response

Cultural characteristics observed after an incubation at different temperatures for 24-48 hours, when subcultured on Brilliant Green Agar Base (M016) and then incubated at 35-37°C for 18-24 hours.

Organism	Inoculum (CFU)	Recovery at 37°C	Recovery at 42 ± 1°C	2 Colour of colony
Escherichia coli ATCC 25922 (00013*)	50-100	fair	poor	yellowish green
<i>Salmonella</i> Paratyphi B ATCC 8759	50-100	good	good	pinkish white
Salmonella Enteritidis ATC 13076 (00030*)	C50-100	luxuriant	luxuriant	pinkish white
Salmonella Typhi ATCC 6539	50-100	fair-good	fair	reddish pink
<i>Salmonella</i> Typhimurium ATCC 14028 (00031*)	50-100	luxuriant	luxuriant	pinkish white

Key : *Corresponding WDCM numbers.

Storage and Shelf Life

6WRUH EHWZHHQ f& LQ D WLJKWO\ FORVHG FRQWDLQHU DQG WKH SU ODEHO 2Q RSHQLQJ SURGXFW VKRXOG EH SURSHUO\ VWRUHG GU\ DIWH GXH WR WKH K\JURVFRSLF QDWXUH RI WKH SURGXFW ,PSURSHU VWRUD, YHQWLODWHG DUHD SURWHFWHG IURP H[WUHPHV RI WHPSHUDWXUH DQG SHUIRUPDQFH LV EHVW LI XVHG ZLWKLQ VWDWHG H[SLU\ SHULRG

Disposal

8VHU PXVW HQVXUH VDIH GLVSRVDO E\ DXWRFODYLQJ DQG RU LQFLQHUD HVWDEOLVKHG ODERUDWRU\ SURFHGXUHV LQ GLVSRVLQJ RI LQIHFWLRXV EH GHFRQWDPLQDWHG DQG GLVSRVHG RI LQ D,FFRUGDQFH ZLWK FXUUHQW

Reference

1. Isenberg, H.D. Clinical Microbiology Procedures Handbook 2nd Edition.

2. Jorgensen, J.H., Pfaller, M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.

3. Rappaport F., Konforti N. and Navon B., 1956, J. Clin. Path., 9:261.

4. Vassiliadis P. Pateraki E., Papaiconomou N., Papadaicis J. A., Trichopoulos D., 1976, Annales de Microbiologie (Institute Pasteur), 127B : 195.

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