



## Chlamyospore Agar

M113

### Intended Use:

Recommended for differentiation of *Candida albicans* from other *Candida* species on the basis of chlamyospore formation.

### Composition\*\*

Ingredients	Gms / Litre
Ammonium sulphate	1.000
Monopotassium phosphate	1.000
Biotin	0.000005
Trypan blue	0.100
Purified polysaccharide	20.000
Agar	15.000
Final pH ( at 25°C)	5.1±0.2

\*\*Formula adjusted, standardized to suit performance parameters

### Directions

Suspend 37.1 grams in 1000 ml purified / distilled water. Heat to boiling to dissolve the medium completely. Sterilize by autoclaving at 15 lbs pressure (121°C) for 15 minutes. Cool to 45-50°C. Mix well and pour into sterile Petri plates.

### Principle And Interpretation

*Candida albicans* is a diploid sexual fungus (a form of yeast), and the causative agent of opportunistic oral and vaginal infections in humans (5). *C. albicans* is a commensal of skin, gastrointestinal and genitourinary tract. However, under certain conditions overgrowth of this results into oesopharyngeal candidiasis, vulvovaginal candidiasis and candidemia. Chlamyospores formation is the most differential characteristic of *C. albicans* (5). Chlamyospore Agar was specially designed for the differentiation of *C. albicans* from other species on the basis of chlamyospores formation. It is prepared according to the formula of Nickerson and Mankowshi (4).

Ammonium sulphate acts as sources of ions that simulate metabolism. Monopotassium phosphate provides buffering to the medium. Biotin provides the necessary vitamins required for metabolism. Purified polysaccharide acts as a source of carbon. Trypan blue is a vital dye absorbed selectively by the chlamyospores and imparts blue colour to chlamyospores, whereas the filaments are colourless.

Test for chlamyospores: Scratch cut mark like X onto the agar surface with inoculum using sterile needle. Aseptically place an alcohol-flamed and cooled cover slip onto the agar surface over the intersecting lines of the cut marks of X. Incubate plates at 20-25°C for 2-6 days. Temperature should not be higher than 25°C since it will not permit chlamyospore formation. Observe the plates under low power of microscope. After incubation, most strains of *C. albicans* and *C. stellatoide* will form typical chlamyospores. Chlamyospores will be seen along the edge of the cover slip. Chlamyospores are round, thick walled, blue coloured and at the terminal ends of hyphae.

Some *C. albicans* strains may lose their ability to produce chlamyospores after repeated subculturing.

### Type of specimen

Food samples; Water samples

### Specimen Collection and Handling

For food samples, follow appropriate techniques for sample collection and processing as per guidelines (1).

For water samples, follow appropriate techniques for sample collection, processing as per guidelines and local standards.(6)

After use, contaminated materials must be sterilized by autoclaving before discarding.

## Warning and Precautions :

Read the label before opening the container. Wear protective gloves/protective clothing/eye protection/ face protection. Follow good microbiological lab practices while handling specimens and culture. Standard precautions as per established guidelines should be followed while handling specimens. Safety guidelines may be referred in individual safety data sheets.

## Limitations :

1. A temperature of 25°C is recommended for the foremost results.

## Performance and Evaluation

Performance of the medium is expected when used as per the direction on the label within the expiry period when stored at recommended temperature.

## Quality Control

### Appearance

Cream to blue homogeneous free flowing powder

### Gelling

Firm, comparable with 1.5% Agar gel

### Colour and Clarity of prepared medium

Blue coloured opaque gel forms in Petri plates

### Reaction

Reaction of 3.71% w/v aqueous solution at 25°C. pH : 5.1±0.2

### pH

4.90-5.30

### Cultural Response

Cultural characteristics observed after an incubation at 20-25°C for 2-6 days.

Organism	Growth	Chlamydo spores
<i>Candida albicans</i> ATCC 10231 (00054*)	good-luxuriant	positive
<i>Candida kruisei</i> ATCC 24408	good-luxuriant	negative
<i>Candida minosa</i>	good-luxuriant	negative
<i>Candida tropicalis</i> ATCC 1369	good-luxuriant	negative

Key : \*Corresponding WDCM numbers.

## Storage and Shelf Life

Store between 10-30°C in a tightly closed container and the prepared medium at 20-30°C. Use before expiry date on the label. On opening, product should be properly stored dry, after tightly capping the bottle in order to prevent lump formation due to the hygroscopic nature of the product. Improper storage of the product may lead to lump formation. Store in dry ventilated area protected from extremes of temperature and sources of ignition Seal the container tightly after use. Use before expiry date on the label.

Product performance is best if used within stated expiry period.

## Disposal

User must ensure safe disposal by autoclaving and/or incineration of used or unusable preparations of this product. Follow established laboratory procedures in disposing of infectious materials and material that comes into contact with sample must be decontaminated and disposed of in accordance with current laboratory techniques (2,3).

## Reference

1. Baird R.B., Eaton A.D., and Rice E.W., (Eds.), 2015, Standard Methods for the Examination of Water and Wastewater, 23rd ed., APHA, Washington, D.C.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook 2nd Edition.
3. Jorgensen, J.H., Pfaller, M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.

4. Nickerson, 1953, J. Infect. Dis., 92:20
5. Ryan K. J., Ray C. G., (Eds.), 2004, Sherris Medical Microbiology, 4th Ed., McGraw Hill.
6. Salfinger Y., and Tortorello M.L., 2015, Compendium of Methods for the Microbiological Examination of Foods, 5th Ed., American Public Health Association, Washington, D.C.

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**Disclaimer :**

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