

# **Technical Data**

## Charcoal Agar Base w/ Niacin

## **Intended Use:**

Recommended for cultivation of Bordetella pertussis and Haemophilus infuenzae.

Composition**	
Ingredients	g / L
Gelatin peptone	10.000
HM peptone B #	10.000
Sodium chloride	5.000
Starch, soluble	10.000
Nicotinic acid (Niacin)	0.001
Charcoal	4.000
Agar	12.000
Final pH ( at 25°C)	$7.4{\pm}0.2$
**Formula adjusted, standardized to suit performance parameters	

<sup>#-</sup>Equivalent to Beef extract

## **Directions**

Suspend 51.0 grams in 900 ml purified/distilled water. Heat to boiling to dissolve the medium completely. Sterilize by autoclaving at 15 lbs pressure (121°C) for 15 minutes. Cool to 45-50°C and aseptically add sterile 10% of defibrinated blood and rehydrated contents of one vial of Bos Selective Supplement (FD004). Mix well and pour into sterile Petri plates. For *Haemophilus* species the medium can be converted to chocolate agar.

## **Principle And Interpretation**

The genus *Bordetella* contains four species : *Bordetella pertussis*, *Bordetella parapertussis*, *Bordetella bronchiseptica* and *Bordetella avium* (1). Genetic studies have shown that these organisms are very closely related to each other. Humans are the only host of *B.pertussis* and *B.parapertussis*, while *B.bronchiseptica* is found in a wide variety of animals and occasionally found in humans (2). *B. avium* is found in birds. *Bordetella* species are obligately aerobic and metabolically not very active. They are non-motile except *B.bronchiseptica*. *B.pertussis* is the major cause of whooping cough or pertussis. *B.parapertussis* is associated with a milder form of the disease (3). Primary isolation of *B.pertussis* in particular, requires the addition of charcoal, 15-20% blood to neutralize the growth-inhibiting effects. Isolation of this organism requires enrichment medium.

Charcoal Agar is prepared according to the method of Mishulow, Sharpe and Cohen (2). This medium can be used as a replacement for Bordet-Gengou Agar for isolation of *B.pertussis* and for the production of *B.pertussis* vaccines. Charcoal Agar supplemented with horse blood can also be used for the cultivation and isolation of *Haemophilus influenzae* (4). Medium ingredients like gelatin peptone and HM peptone B provide nitrogen and carbon compounds, long chain amino acids and other essential nutrients to the organisms. Sodium chloride maintains osmotic balance. Starch soluble and charcoal neutralizes substances toxic to *Bordetella* species such as fatty acids. Charcoal has the tendency to settle at the bottom of the flask. Therefore, before dispensing, swirl the flasks gently to obtain a uniform charcoal suspension (5). The difficulty in the isolation of *Bordetella pertussis* from nasopharyngeal secretions is the repression of unwanted flora during the long incubation period on nutritious media. Penicillin can be added to the medium as an antimicrobial agent for restricting the other contaminants. However Penicillin resistant flora still causes the contamination that was observed by Lacey (4). Necessity of the Nicotinic acid as a growth factor was showed by Proom (6). Methicillin was found to be superior to Penicillin in suppressing unwanted nasopharyngeal flora as observed by Broome et al (7). Sutcliffe and Abbott found that Cephalexin was still better than Methicillin (8).

The medium can also be used for the maintenance of stock cultures of *Bordetella pertussis* on slants with weekly subcultures. Charcoal Agar with Niacin can be converted to Chocolate Agar for isolation of *Haemophilus* species.

## Type of specimen

Clinical samples : Sputum, cerebrospinal fluid

## **Specimen Collection and Handling**

For clinical samples follow appropriate techniques for handling specimens as per established guidelines (9,10). After use, contaminated materials must be sterilized by autoclaving before discarding.

#### Warning and Precautions

In Vitro diagnostic use only. For professional use only. Read the label before opening the container. Wear protective gloves/protective clothing/eye protection/face protection. Follow good microbiological lab practices while handling specimens and culture. Standard precautions as per established guidelines should be followed while handling clinical specimens. Safety guidelines may be referred in individual safety data sheets.

#### Limitations

1. Some Haemophilus species will grow on Bordetella isolation media and cross-react with B. pertussis antisera.

2. *B. pertussis* colonies may not be visible without the aid of a microscope after 2-4 days.

#### **Performance and Evaluation**

Performance of the medium is expected when used as per the direction on the label within the expiry period when stored at recommended temperature.

## **Quality Control**

## Appearance

Grey to greyish black homogeneous free flowing powder

#### Gelling

Firm, comparable with 1.2% Agar gel

#### Colour and Clarity of prepared medium

Black coloured, opaque gel with undissolved black particles forms in Petri plates

#### Reaction

Reaction of 5.1% w/v aqueous solution at 25°C. pH : 7.4±0.2

## pН

#### 7.20-7.60

#### **Cultural Response**

Cultural characteristics observed with added sterile defibrinated blood and Bos Selective Supplement (FD004), after an incubation at 35 - 37°C for 24 - 48 hours.

Organism	Inoculum (CFU)	Growth	Recovery
Bordetella bronchiseptica ATCC 4617	50-100	good-luxuriant	>=50%
Bordetella parapertussis ATCC 15311	50-100	good-luxuriant	>=50%
Bordetella pertussis ATCC 8467	50-100	good-luxuriant	>=50%
Staphylococcus aureus subsp. aureus ATCC 25923 (00034*)	>=10 <sup>4</sup>	inhibited	0%
Klebsiella pneumoniae ATCC 13883 (00097*)	>=10 <sup>4</sup>	inhibited	0%

Key: (\*) Corresponding WDCM numbers.

## **Storage and Shelf Life**

Store between 10-30°C in a tightly closed container and the prepared medium at 2-8°C. Use before expiry date on the label. On opening, product should be properly stored dry, after tightly capping the bottle in order to prevent lump formation due to the hygroscopic nature of the product. Improper storage of the product may lead to lump formation. Store in dry ventilated area protected from extremes of temperature and sources of ignition Seal the container tightly after use. Product performance is best if used within stated expiry period.

#### **Disposal**

User must ensure safe disposal by autoclaving and/or incineration of used or unusable preparations of this product. Follow established laboratory procedures in disposing of infectious materials and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (9,10).

#### Reference

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