Sterile Saline, 0.9%

LQ199D

Intended Use:

Sterile, ready prepared saline used as a diluent

Composition**

Ingredients	g/ L
Sodium chloride	9.000

**Formula adjusted, standardized to suit performance parameters

Directions

Label the ready to use LQ199D bottle. Inoculate the bottle with the pre-determined volume of the sample or 50-100 CFU of a known culture (as positive control) and incubate at 35 - 37°C for 24-48 hours.

Principle And Interpretation

Saline solution maintains the osmotic balance in microbial cells and helps to maintain the cell integrity and viability. Normal saline is used for preparing microbial suspensions for detection of antimicrobial agents, or to growth media used for disk susceptibility testing. It is also used in the preparing of stock solutions and serial dilutions of antimicrobial agents.

Type of specimen

Serial dilution of samples

Specimen Collection and Handling

For serial dilution samples follow appropriate techniques for handling specimens as per established guidelines.
After use, contaminated materials must be sterilized by autoclaving before discarding.

Warning and Precautions

Read the label before opening the container. Wear protective gloves/protective clothing/eye protection/face protection. Follow good microbiological lab practices while handling specimens and culture. Standard precautions as per established guidelines should be followed while handling specimens. Safety guidelines may be referred in individual safety data sheets.

Limitations

1. For preparing dilution of spore suspension addition of surfactant is required.

Performance and Evaluation

Performance of the medium is expected when used as per the direction on the label within the expiry period when stored at recommended temperature.

Quality Control

Appearance

Sterile clear Saline 0.9% Solution in glass bottle.

Colour

Colourless solution.

Quantity of medium

500 ml of medium in glass bottle

Sterility Check

Passes release criteria

Cultural Response

Cultural characteristics observed after recovery on Soybean Casein Digest Agar after an incubation at 30-35°C for 18-24 hours for bacteria and Sabouraud Dextrose Agar at 30-35°C for 24-48 hours.

Organism	Inoculum (CFU)	Recovery within 2 hours of incubation	Recovery within 4 hours of incubation
Escherichia coli ATCC 8739 (00012*)	50 -100	no decrease in colony count	no decrease in colony count
Escherichia coli ATCC 25922 (00013*)	50 -100	no decrease in colony count	no decrease in colony count
Staphylococcus aureus subsp. *aureus* ATCC 6538 (00032*)	50 -100	no decrease in colony count	no decrease in colony count
^*Pseudomonas paraeruginosa ATCC 9027 (00026*)*	50 -100	no decrease in colony count	no decrease in colony count
Pseudomonas aeruginosa ATCC 27853 (00025)*	50 -100	no decrease in colony count	no decrease in colony count
Salmonella Typhimurium ATCC 14028 (00031)*	50 -100	no decrease in colony count	no decrease in colony count
Salmonella Abony NCTC 6017 (00029*)	50 -100	no decrease in colony count	no decrease in colony count
#*Bacillus spizizennii* ATCC 6633 (00003*)	50 -100	no decrease in colony count	no decrease in colony count
$ *Kocuria rhizophila* ATCC 9341	50 -100	no decrease in colony count	no decrease in colony count
Candida albicans ATCC 10231 (00054*)	50 -100	no decrease in colony count	no decrease in colony count
Candida albicans ATCC 2091	50 -100	no decrease in colony count	no decrease in colony count

Key : (*) Corresponding WDCM numbers,

^ Formerly known as *Pseudomonas aeruginosa*

Formerly known as *Bacillus subtilis* subsp. *spizizenii*

$ Formerly known as *Micrococcus luteus*

Storage and Shelf Life

On receipt store between 15-30°C. Use before expiry date on the label. Product performance is best if used within stated expiry period.

Disposal

User must ensure safe disposal by autoclaving and/or incineration of used or unusable preparations of this product. Follow established laboratory procedures in disposing of infectious materials and material that comes into contact with sample must be decontaminated and disposed of in accordance with current laboratory techniques (1,2).

Reference

1. Isenberg, H.D. Clinical Microbiology Procedures Handbook 2nd Edition.

2. Jorgensen, J.H., Pfaller, M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.

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