

Nitrate Broth

LQ083

Intended Use:

Sterile, ready prepared medium for detection of nitrate reduction by bacteria.

Composition**

Ingredients	g / L
Peptone	5.000
HM peptone B #	3.000
Potassium nitrate	1.000
Final pH (at 25°C)	7.0±0.2

**Formula adjusted, standardized to suit performance parameters

Equivalent to Beef extract

Directions

Label the ready to use LQ083 bottle. Inoculate 50-100 cfu sample and Incubate at specified temperature and time.

Principle And Interpretation

Nitrate Broth is prepared in accordance with the formula published in Pure Culture Study of Bacteria of the Society of American Bacteriologist (1). The ability to reduce nitrate is valuable for differentiating and identifying various types of bacteria especially *Enterobacteriaceae* family (2). Non-fermenters and other miscellaneous gram-negative bacilli vary in their ability to reduce nitrates. Some members of this group are capable of denitrification which is the reduction of nitrate to nitrogen gas. For the glucose fermenting gram-negative bacilli, the production of nitrogen gas from nitrate is an important differential test (3).

Potassium nitrate in the medium acts as a substrate for determining nitrate reduction by bacteria. Certain bacteria convert nitrate to nitrite, ammonia or nitrogen gas. The presence of nitrites can be detected by the addition of 0.5 ml each of sulphanilic acid (R015) and alpha-naphthylamine solution (R009). The development of red violet colour, due to the formation of a red diazonium dye i.e. p-sulfobenzene-azo-a-naphthylamine, indicates nitrate reduction to nitrite. If no colour develops, it means that either nitrate is not reduced or further reduction to ammonia or nitrogen gas has taken place. This can be verified by adding a pinch of zinc dust to the tube. Zinc reduces nitrate to nitrite resulting in a red colour. The red colour indicates that nitrate is still present and was not reduced previously. An absence of red colour after the addition of zinc dust indicates that no nitrate is present and thus the nitrate was reduced further than nitrite. Therefore the nitrate reduction test is evidenced by either the presence of a catabolic end product or the absence of nitrate in the medium. Members of *Enterobacteriaceae* characteristically reduce nitrate to nitrite which reacts with sulphanilic acid and N, N-dimethyl-1-naphthylamine to produce the red colour. This reaction is known as Griess reaction. If an organism grows rapidly and reduces nitrate actively, the test should be performed after an early incubation period since the nitrite may be further reduced to nitrogen.

Type of specimen

Isolated Microorganisms from clinical and non-clinical samples

Specimen Collection and Handling:

For the test: Add few drops of each reagent i.e. sulphanilic acid (R015) and a-naphthylamine solution (R009) into the tube containing culture to be tested. A distinct red or pink colour indicates nitrate reduction. A control (un-inoculated) tube should also be tested. If there is no pink colour formation, add a pinch of zinc dust to confirm the absence of nitrate in the medium (3).

Warning and Precautions :

In Vitro diagnostic use. For professional use only. Read the label before opening the container. Wear protective gloves/ protective clothing/eye protection/face protection. Follow good microbiological lab practices while handling specimens and culture. Standard precautions as per established guidelines should be followed while handling specimens. Safety guidelines may be referred in individual safety data sheets.

Limitations :

1. Nitrate reduction is not a confirmatory test. Complete identification of bacteria should include the morphology, gram reaction, biochemical and serological tests. Addition of excess zinc may result in false negative reaction. Also during performance of nitrate reduction test with α -naphthylamine, the colour produced in a positive reaction may fade quickly (4).
2. The nitrate test is very sensitive and an uninoculated nitrate control should be tested with reagents to determine whether medium is nitrite free.(4)

Performance and Evaluation

Performance of the medium is expected when used as per the direction on the label within the expiry period when stored at recommended temperature.

Quality Control

Appearance

Sterile clear Nitrate Broth in bottles.

Colour

Light amber coloured solution.

Quantity of Medium

5 ml of medium in bottle.

Sterility Check

Passes release criteria

pH

6.80-7.20

Cultural Response

Cultural characteristics observed after an incubation at 35-37°C for 18-24 hours.

Organism	Growth	Nitrate reduction
<i>Acinetobacter calcoaceticus</i> ATCC 19606	luxuriant	negative reaction
<i>Escherichia coli</i> ATCC 25922 (00013*)	luxuriant	Positive reaction, distinct red-pink colour, pink colour developed within 1-2 minutes
# <i>Klebsiella aerogenes</i> ATCC 13048 (00175*)	luxuriant	Positive reaction, distinct red-pink colour, pink colour developed within 1-2 minutes
<i>Salmonella</i> Typhimurium ATCC 14028 (00031*)	luxuriant	Positive reaction, distinct red-pink colour, pink colour developed within 1-2 minutes

Key : (*) Corresponding WDCM numbers. (#) Formerly known as *Enterobacter aerogenes*

Storage and Shelf Life

On receipt store between 15-30°C. Use before expiry date on the label Product performance is best if used within stated expiry period.

Disposal

User must ensure safe disposal by autoclaving and/or incineration of used or unusable preparations of this product. Follow established laboratory procedures in disposing of infectious materials and material that comes into contact with sample must be decontaminated and disposed of in accordance with current laboratory techniques (5,6).

Reference

1. Society of American Bacteriologist, Pure Culture Study of Bacteria, 1944, 12 : Leaflet 11: 8.
2. Ewing, 1986, Edwards and Ewings Identification of *Enterobacteriaceae*, 4th ed., Elsevier Science Pub. Co., Inc., N.Y.
3. MacFaddin, 2000, Biochemical Tests for the Identification of Medical Bacteria, 3rd ed., Williams and Wilkins, Baltimore.
4. MacFaddin J. F., 1985, Media for Isolation-Cultivation-Identification-Maintenance of Medical Bacteria, Vol. 1, Williams and Wilkins, Baltimore.

5. Isenberg, H.D. Clinical Microbiology Procedures Handbook 2nd Edition.
6. Jorgensen, J.H., Pfaller, M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.

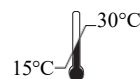
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