

Technical Data

HiCombiTM Dual Performance Medium

Intended use

Recommended for rapid growth of *Enterobacteria, Pseudomonas*, Staphylococci and *Candida*. Combination of solid (20ml) and liquid (40ml) media in single bottle.

Composition**

Ingredients	g / L
Solid	20.000 ml
HM infusion powder #	12.500
BHI powder	5.000
Proteose peptone	10.000
Dextrose (Glucose)	2.000
Sodium chloride	5.000
Disodium hydrogen phosphate	2.500
Agar	15.000
Liquid	40.000 ml
Same as solid media without Agar	

**Formula adjusted, standardized to suit performance parameters

Equivalent to Calf brain infusion from

Directions

Label the ready to use blood culture bottle. Do not unscrew the cap. Remove the top seal of the cap. Disinfect the part of the rubber stopper which is now exposed. Draw patient's blood with the sterile or disposable needle and syringe. Transfer the blood sample immediately into the culture bottle by puncturing the rubber stopper with the needle and injecting the blood. Incubate the bottle for 4-6 hours at 30 -35°C. For adsorption on solid surface. DO NOT SHAKE OR HOLD MORE THAN 15 SECONDS. Revert into an upright position and incubate for 18-24 hours at 30-35°C or longer if necessary. Venting: Use sterile venting needle (LA038). Keep the bottle in an upright position preferably in a biological safety cabinet, place an alcohol swab over the rubber stopper and insert the venting needle with filter through it. Insertion and withdrawal of the needle should be done in a straight line. discard the needle and mix the contents by gently inverting the bottle 2-3 times. Do Not vent the bottle for anaerobic cultures. Incubate at 30-35°C for 18-24 hours and further for seven days. Recommended volume of blood to be tested in LQ012: 8-10 ml (For Adult use).

Principle And Interpretation

BHI Medium is useful for cultivating a wide variety of microorganisms since it is a highly nutritive medium. It is also used to prepare the inocula for antimicrobial susceptibility testing. BHI Broth is a modification of the original formulation of Rosenow, where he added pieces of brain tissues to dextrose broth (1). BHI Broth is also the preferred medium for anaerobic bacteria, yeasts and moulds (2,3). This medium is nutritious and well buffered to support the growth of wide variety of organisms (1,4,5). With the addition of 10% defibrinated sheep blood, it is useful for isolation and cultivation of *Histoplasma capsulatum* (6) and other fungi. For selective isolation of fungi, addition of gentamicin and/or chloramphenicol is recommended (7).

Proteose peptone, HM infusion powder and BHI powder serve as sources of carbon, nitrogen, essential growth factors, amino acids and vitamins. Dextrose serves as a source of energy. Disodium phosphate helps in maintaining the buffering action of the medium whereas sodium chloride maintains the osmotic equilibrium of the medium.

Type of specimen

Clinical samples : Blood

Specimen Collection and Handling

For clinical samples follow appropriate techniques for handling specimens as per established guidelines (8,9). After use, contaminated materials must be sterilized by autoclaving before discarding.

Warning and Precautions

In Vitro diagnostic use only. For professional use only. Read the label before opening the container. Wear protective gloves/protective clothing/eye protection/face protection. Follow good microbiological lab practices while handling

specimens and culture. Standard precautions as per established guidelines should be followed while handling clinical specimens. Safety guidelines may be referred in individual safety data sheets.

Limitations

1. Further biochemical and serological testing is required for complete identification.

2. Individual organisms differ in their growth requirement and may show variable growth patterns on the medium.

3. Each lot of the medium has been tested for the organisms specified on the COA. It is recommended to users to validate the medium for any specific microorganism other than mentioned in the COA based on the user's unique requirement.

Performance and Evaluation

Performance of the medium is expected when used as per the direction on the label within the expiry period when stored at recommended temperature.

Quality Control

Appearance

In a sterile glass bottle combination of broth and one agar coated surface.

Colour of Agar medium	Colour of liquid medium

Yellow coloured media	Amber coloured solution
Quantity of medium	
20ml of medium in glass b	ottle 40ml of medium in glass bottle
pH of Agar medium	pH of liquid medium
7.20-7.60	7.20- 7.60

Sterility Check

Passes release criteria

Cultural response

Cultural characteristics was observed after incubation at 35-37°C for 18-48 hours.

Organism	Inoculum (CFU)	Growth on agar medium	Growth on liquid medium
Candida albicans ATCC 10231 (00054*)	50-100	Luxuriant	Luxuriant
Haemophilus influenzae ATCC 19418	50-100	Luxuriant	Luxuriant
Pseudomonas aeruginosa ATCC 27853 (00025*)	50-100	Luxuriant	Luxuriant
Streptococcus pyogenes ATCC 19615	50-100	Luxuriant	Luxuriant
Staphylococcus aureus subsp. aureus ATCC	50-100	Luxuriant	Luxuriant
25923 (00034*)			
Neisseria meningitidis ATCC 13090	50-100	Luxuriant	Luxuriant
Streptococcus pneumoniae ATCC 6303	50-100	Luxuriant	Luxuriant

Key : (*) Corresponding WDCM numbers.

Storage and Shelf Life

On receipt store between 15-30°C. Use before expiry date on the label. Product performance is best if used within stated expiry period.

Disposal

User must ensure safe disposal by autoclaving and/or incineration of used or unusable preparations of this product. Follow established laboratory procedures in disposing of infectious materials and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (8,9).

Reference

1.Rosenow, 1919, J. Dental Research, 1:205.

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3. Atlas R. M., 1993, Handbook of Microbiological Media, 147-153, CRC Press, Boca Raton, FL.

4.Roseburg T. et al, 1944, J. Inf. Dis., 74:

5. Conant N. F., 1950, Diagnostic Procedures and Reagents, 3rd Ed., APHA Inc., New York

6.Howard B., Keiser J. F., Weissfeld A. et al, 1994, Clinical and Pathogenic Microbiology, 2nd Ed., Mosby Co.

7.Murray P. R., Baron J. H., Pfaller M. A., Jorgensen J. H. and Yolken R. H., (Eds.), 2003, Manual of Clinical Microbiology,8th Ed., American Society for Microbiology, Washington, D.C.

8. Isenberg, H.D. Clinical Microbiology Procedures Handbook 2nd Edition.

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Revision : 03/2023



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