



Ceftazidime/ Ceftazidime + Clavulanic acid Ezy MIC™ Strip EM098I (CAZ/CAZ+)

(Ceftazidime (CAZ): 0.5-32)

(Ceftazidime + Clavulanic acid (CAZ+): 0.064-4)

(Individually Packed)

Antimicrobial Susceptibility Testing

For *In Vitro* Diagnostic use

It is a Phenotypic ESBL detection strip which is coated with Ceftazidime with & without clavulanic acid on a single strip in a concentration gradient manner. The upper half has Ceftazidime + Clavulanic acid with highest concentration tapering downwards, whereas lower half is similarly coated with Ceftazidime in a concentration gradient in reverse direction

Introduction:

Ezy MIC™ strip is useful for quantitative determination of susceptibility of bacteria to antibacterial agents. The system comprises of a predefined quantitative gradient which is used to determine the Minimum Inhibitory Concentration (MIC) in mcg/ml of different antimicrobial agents against microorganisms as tested on appropriate agar media, following overnight incubation.

Ezy MIC™ Strip FEATURES AND ADVANTAGES

Ezy MIC™ strip exhibits several advantages over existing plastic strip.

1. Ezy MIC™ strip is made up of porous paper material unlike plastic non-porous material
2. Ezy MIC™ strip has MIC values printed on both sides identically.
3. The antimicrobial agent is evenly distributed on either side of the Ezy MIC™ strip and hence it can be placed by any side on the agar surface.
4. For Ezy MIC™ strips, MIC values can be read without opening the lid of the plate as most commonly translucent medium such as Mueller Hinton Agar is employed.
5. Once placed, Ezy MIC™ strip is adsorbed within 60 seconds and firmly adheres to the agar surface.
6. Unlike the plastic material, it does not form air bubbles underneath and hence there is no need to press the strip once placed.

Principle and Interpretation

ESBLs are enzymes that mediate resistance to extended-spectrum (third generation) cephalosporins (e.g., Cefotaxime, Ceftazidime, and Ceftriaxone) and monobactams (e.g., Aztreonam) but do not affect cephamycins (e.g., Cefoxitin and Cefotetan) or carbapenems (e.g., Meropenem or Imipenem). The presence of an ESBL-producing organism in a clinical infection can result in treatment failure if one of the above classes of drugs is used. ESBLs can be difficult to detect because they have different levels of activity against various cephalosporins. Thus, the choice of which antimicrobial agents to test is critical. If an ESBL is detected, all penicillins, cephalosporins, and Aztreonam should be reported as resistant, even if *in vitro* test results indicate susceptibility.

CLSI has developed initial micro dilution screening tests using selected antimicrobial agents (1). Following antimicrobial concentration is to be used for initial screen test:

For *K. pneumoniae*, *K. oxytoca* and *E. coli* use Cefpodoxime 4 µg/ml or Ceftazidime 1 µg/ml or Aztreonam 1 µg/ml or Cefotaxime 1 µg/ml or Ceftriaxone 1 µg/ml and for *P. mirabilis* use Cefpodoxime 1 µg/ml or Ceftazidime 1 µg/ml or Cefotaxime 1 µg/ml (The use of more than one antimicrobial agent for screening improves sensitivity of detection).

Growth at or above the screening concentrations may indicate ESBL production i.e. for *K. pneumoniae*, *K. oxytoca* and *E. coli* MIC of ≥ 8 µg/ml for Cefpodoxime and ≥ 2 µg/ml for Ceftazidime, Aztreonam, Cefotaxime and Ceftriaxone while a MIC of ≥ 2 µg/ml for Cefpodoxime, Ceftazidime and Cefotaxime for *P. mirabilis* is indicative of ESBL production.

Strains of *Klebsiella* spp. and *Escherichia coli* that produce ESBLs may be clinically resistant to therapy with penicillins, cephalosporins, or Aztreonam, despite apparent *in vitro* susceptibility to some of these agents.

CLSI recommends performing phenotypic confirmation of potential ESBL-producing isolates by testing both Cefotaxime and Ceftazidime, alone and in combination with Clavulanic acid (1). Testing can be performed by the broth micro dilution method or by disk diffusion. *K. pneumoniae* ATCC 700603 and *E. coli* ATCC 25922 should be used for quality control of ESBL tests (1).

For dilution method; **the culture is to be reported as ESBL positive if a ≥ 3 twofold concentration decrease in MIC for antibiotic tested in combination with clavulanic acid vs its MIC when tested alone is obtained.**

However, the phenotypic confirmatory test does not detect all ESBLs. Some organisms with ESBLs contain other β -lactamases that can mask ESBL production in the phenotypic test, resulting in a false-negative test. Currently, detection of organisms with multiple β -lactamases that may interfere with the phenotypic confirmatory test can only be accomplished using isoelectric focusing and DNA sequencing.

If an isolate is confirmed as an ESBL-producer by the CLSI-recommended phenotypic confirmatory test procedure, all penicillins, cephalosporins, and Aztreonam should be reported as resistant. This list does not include the cephamycins (Cefotetan and Cefoxitin), which should be reported according to their routine test results. If an isolate is not confirmed as an ESBL-producer, current recommendations suggest reporting results as for routine testing. Do not change interpretations of penicillins, cephalosporins, and Aztreonam for isolates not confirmed as ESBLs.

Other isolates of *Enterobacteriaceae*, such as *Salmonella* species and *P. mirabilis*, and isolates of *P.aeruginosa* produce ESBLs. Though screening of *P. mirabilis* for ESBL production is recommended only when it is deemed clinically relevant (e.g. bacteremic isolate). The decision to perform ESBL screening tests to all urine isolates should be made on an institutional basis, considering prevalence, therapy, and infection control issues.

METHOD AND USE OF EZY MIC™ STRIPS

- **Type of specimen**

Pure cultures should be derived from specimens obtained from patients prior to the initiation of antimicrobial therapy. Specimens can be of bacterial or fungal isolates derived from blood, urine, faeces, pus, CSF etc. Direct specimens should not be employed in this test. Refer procedure, which includes preparation of inoculum (1, 3).

- **Clinical specimen collection, handling and processing**

Follow appropriate techniques for handling specimens as per established guidelines. After use, contaminated materials must be sterilized by autoclaving before discarding (1, 3).

- **Guidelines for preparation of the medium**

Prepare the medium of choice from dehydrated powder according to the directions specified on the label. Cool the sterilized molten medium to 45-50°C and pour in sterile, dry Petri plates on a leveled surface, to a depth of 4 ± 0.2 mm and allow solidifying. Few droplets appearing on the surface of the medium following cooling do not matter. Hence, once poured, Petri plates containing media should not be dried on laminar flow and can be used immediately for swabbing.

- **Preparation of Inoculum**

Use only pure cultures. Confirm by Gram-staining before starting susceptibility test. Transfer 4-5 similar colonies with a wire, needle or loop to 5 ml Tryptone Soya Broth (M011) and incubate at 35-37°C for 2-8 hours until light to moderate turbidity develops. Compare the inoculum turbidity with that of standard 0.5 McFarland. Alternatively, the inoculum can be standardized by other appropriate optical method (0.08 - 0.13 OD turbid suspension at 620 nm). Also direct colony suspension method can be used. Prepare a direct colony suspension, from 18-24 hour old non-selective media agar plate in broth or saline. Adjust the turbidity to that of standard 0.5 McFarland. This method is recommended for testing fastidious organisms like *Haemophilus* spp., *Neisseria* spp, and streptococci and for testing staphylococci for potential Methicillin or Oxacillin resistance.

- **Test Procedure**

1. Prepare plates with suitable make of recommended medium as mentioned in Table 2 & Table 4.
2. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Swab the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking.
3. Remove individually packed Ezy MIC™ strip from cold and keep it at room temperature for 15 minutes before opening.
4. Tear the aluminum pouch carefully at the notch provided, so that it does not damage the Ezy MIC™ Strip.
5. With the help of sterile forceps, remove the Ezy MIC™ Strip gently and place the strip at a desired position on an agar plate swabbed with the test culture.
6. DO NOT PRESS Ezy MIC™ STRIP. Within 60 seconds, Ezy MIC™ strip will be adsorbed and will firmly adhere to the agar surface.
7. Ezy MIC™ strip should not be repositioned or adjusted once placed.
8. Transfer plates in the incubator under appropriate conditions.

Reading of IC (Inhibitory Concentration) values:

1. Read the plates only when sufficient growth is seen.
2. Read the MIC where the ellipse intersects the MIC scale on the strip.
3. For bactericidal drugs such as members of β-lactams class of drugs, Amikacin, Vancomycin, Gentamicin always read the MIC at the point of completion inhibition of all growth, including hazes, microcolonies and isolated colonies. If necessary, use magnifying glass.
4. Isolated colonies, microcolonies and hazes appearing in the zone of inhibition are indicative of hetero nature of the culture having resistant subpopulation in it. In such cases, consider reading for IC determination at a point on the scale above which no resistant colonies are observed close to strip (within 1-3 mm distance from the strip).
5. If the ellipse intersects the strip in between 2 dilutions, read the IC value which is nearest to the intersection

Warning and Precautions:

1. Ezy MIC™ Strip is intended for *In vitro* diagnostic use only.
2. Although based on simple procedure, Ezy MIC™ Strip should only be used by at least semi-trained personnel.
3. This strip is intended only for agar diffusion method and not for broth dilution method.
4. Ezy MIC™ Strip should be used strictly according to procedures described herein.
5. Performance of Ezy MIC™ Strips depends on use of proper inoculum and control cultures, recommended test medium and proper storage temperature.
6. Follow aseptic techniques and precautions against microbiological hazards should be used when handling bacterial or fungal specimen throughout the testing procedure.
7. Before using Ezy MIC™ Strips, ensure that the strips are at room temperature.
8. When applying strips be steady. Do not move the strip once in contact with agar surface, since the antibiotic instantaneously diffuse on contact with agar.
9. Place the unused strips back to recommended temperature.

INTERPRETATION & QUALITY CONTROL (As per CLSI Guidelines) :
Interpretation:

Table 1: Use following interpretive criteria for susceptibility categorization.

Report	Formula	Interpretative Criteria
ESBL positive strain	$\frac{CAZ}{CAZ+} = > 8$	When the ratio of the value obtained for Ceftazidime (CAZ) : the value of Ceftazidime in combination with Clavulanic acid (CAZ+) is more than 8 or No zone is obtained for CAZ and Zone obtained in CAZ+

ESBL negative strain	$\frac{\text{CAZ}}{\text{CAZ}^+} = \leq 8$	When Ratio of the value obtained for Ceftazidime (CAZ): the value of Ceftazidime in combination with Clavulanic acid (CAZ+) is less than or equal 8.
ESBL (non-conclusive)		When no zone of inhibition is obtained on either side. In such cases resistance may be due to mechanisms other than ESBL production. These have to be further investigated before reporting.

Quality control

Table 2: Quality control of Ezy MIC™ Strip is carried out by testing the strips with standard ATCC Cultures recommended by CLSI on suitable medium incubated appropriately.

Organism	Medium used	Incubation	Standard
<i>K. pneumoniae</i> ATCC 700603	Mueller Hinton Agar	35-37°C for 18 hrs.	When the ratio of the value obtained for Ceftazidime (CAZ) : the value of Ceftazidime in combination with Clavulanic acid (CAZ+) is more than 8
<i>E. coli</i> ATCC 25922	Mueller Hinton Agar	35-37°C for 18 hrs.	When Ratio of the value obtained for Ceftazidime (CAZ): the value of Ceftazidime in combination with Clavulanic acid (CAZ+) is less than or equal 8.

Storage & Shelf Life:

1. Once the consignment is received, store applicators at Room Temperature and Ezy MIC™ strips container at -20°C or below.
2. Use before expiry date on the label.
3. Ezy MIC™ Strip left over from opened package must be kept dry.
4. Moisture should be prevented from penetrating into or forming within the package or storage container.
5. Check whether the batch number and expiry date are marked on the storage container.
6. Product performance is best within stated expiry period if correctly stored and handled.

Disposal:

After use, Ezy MIC™ Strips and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (2, 3).

Limitation of Test

Ezy MIC™ Strips provides *in vitro* MIC values, which provides only a possible insinuation of pathogens potential in *In vivo* susceptibility. These values can be considered as a guide to therapy selection only after taking into consideration several other factors; and must be the sole decision and responsibility of the physician along with the clinical experience in treating the infection. These tests are comparable to the standards as per the given specifications and set of experiment standards as far as possible. Please refer to CLSI standards for detailed limitation of susceptibility test on the clinical use of an antibiotic in various therapeutic conditions.

References:

1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 1, Section 2.
2. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition, Vol. 3, Section 15.
3. Jorgensen, J.H., Pfaller, M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock, D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
4. Performance Standards of Antimicrobial Susceptibility Testing; 34th Edition. M100-Ed34, Vol.44, No.5, Jan-2024.

Packing:

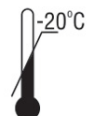
Each Pack contains following material packed in a desiccator with desiccator.

- 1) Ceftazidime/ Ceftazidime + Clavulanic acid Ezy MIC™ Strip (Individually packed) (10/30/60/90/120/150 Strips per pack)
- 2) Package insert

Revision: 02/2024



On receipt store at



In vitro diagnostic
medical device



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Road No. 21Y, MIDC,
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Indicates a single
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system



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