



# Medium 199

With Earle's salts and 25mM HEPES buffer Without L-Glutamine and Sodium bicarbonate

**Product Code: AT094A** 

# **Product Description:**

Medium 199 was the first nutritionally defined medium developed by Morgan, Morton, and Parker in 1950. This complex medium was formulated specifically for nutritional studies on primary chick embryo fibroblasts in the absence of any additives. It was observed that explanted tissue could survive in Medium 199 without serum but long term cultivation of cells required supplementation of the medium with serum.

Medium 199 is formulated with either Hank's salts or Earle's salts. The medium when supplemented with serum can be used for growth of a wide variety of cells. Medium 199 is presently used for the maintenance of non-transformed cells, vaccine and virus production and primary explants of epithelial cells.

AT094A is Medium 199 with Earle's salts, 25mM HEPES buffer. It does not contain L-Glutamine. HEPES, a zwitterionic buffer having a pKa of 7.3 at 37°C prevents the initial rise in pH that tends to occur at the initiation of a culture and increases the buffering capacity of the medium. Users are advised to review the literature for recommendations regarding medium supplementation and physiological growth requirements specific for different cell lines.

## **Composition:**

Composition:		Nicotinamide	0.025
Ingredients	mg/L	Nicotinic acid	0.025
INORGANIC SALTS	8	Pyridoxal hydrochloride	0.025
Calcium chloride dihydrate	265.000	Pyridoxine hydrochloride	0.025
Ferric nitrate nonahydrate	0.720	Retinol Acetate	0.140
Magnesium sulphate anhydrous	97.720	Riboflavin	0.010
Potassium chloride	400.000	Thiamine hydrochloride	0.010
Sodium acetate anhydrous	50.000	i-Inositol	0.050
Sodium chloride	6800.000	p-Amino benzoic acid (PABA)	0.050
Sodium dihydrogen phosphate anhydrous	122.000	OTHERS	
AMINO ACIDS		Adenine sulphate	10.000
Glycine	50.000	Adenosine monophosphate	0.200
L-Alanine	25.000	Adenosine triphosphate	1.000
L-Arginine hydrochloride	70.000	Cholesterol	0.200
L-Aspartic acid	30.000	Deoxyribose	0.500

L-Cysteine hydrochloride monohydrate	0.100
L-Cystine dihydrochloride	26.000
L-Glutamic acid	67.000
L-Histidine hydrochloride monohydrate	22.000
L-Hydroxyproline	10.000
L-Isoleucine	20.000
L-Leucine	60.000
L-Lysine hydrochloride	70.000
L-Methionine	15.000
L-Phenylalanine	25.000
L-Proline	40.000
L-Serine	25.000
L-Threonine	30.000
L-Tryptophan	10.000
L-Tyrosine disodium salt	57.660
L-Valine	25.000
VITAMINS	
Ascorbic acid	0.050
Calciferol	0.100
Choline chloride	0.500
D-Biotin	0.010
D-Ca-Pantothenate	0.010
DL-Tocopherol phosphate disodium salt	0.010
Folic acid	0.010
Menadione	0.010
Nicotinamide	0.025
Nicotinic acid	0.025
Pyridoxal hydrochloride	0.025
Pyridoxine hydrochloride	0.025
Retinol Acetate	0.140
Riboflavin	0.010
Thiamine hydrochloride	0.010
i-Inositol	0.050
p-Amino benzoic acid (PABA)	0.050
OTHERS	
Adenine sulphate	10.000
Adenosine monophosphate	0.200
Adenosine triphosphate	1.000

Glucose	1000.000
Glutathione reduced	0.050
Guanine hydrochloride	0.300
HEPES Buffer	5958.000
Hypoxanthine	0.354
Phenol red	15.000
Polysorbate 80	4.900
Ribose	0.500
Thymine	0.300
Uracil	0.300
Xanthine	0.344

## **Directions:**

- 1. Suspend 15.5gms in 900ml tissue culture grade water with constant, gentle stirring until the powder is completely dissolved. Do not heat the water.
- 2. Add 0.1gms of L-glutamine (TC230) or 3.42ml of 200mM L-glutamine (TCL012) and 2.2gms of sodium bicarbonate powder (TC243) or 29.3ml of 7.5% sodium bicarbonate solution (TCL013) for 1 litre of medium and stir until dissolved.
- 3. Adjust the pH to 0.2 0.3 pH units below the desired pH using 1N HCl or 1N NaOH since the pH tends to rise during filtration.
- 4. Make up the final volume to 1000ml with tissue culture grade water.
- 5. Sterilize the medium immediately by filtering through a sterile membrane filter with a porosity of 0.22 micron or less, using positive pressure rather than vacuum to minimize the loss of carbon dioxide.
- 6. Aseptically add sterile supplements as required and dispense the desired amount of sterile medium into sterile containers.
- 7. Store liquid medium at 2-8°C and in dark till use.

# Material required but not provided:

Tissue culture grade water (TCL010) Sodium bicarbonate (TC230) Sodium bicarbonate solution, 7.5% (TCL013) L-Glutamine powder (TC243) L-Glutamine solution 200mM (TCL012) 1N Hydrochloric acid (TCL003)

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1N Sodium hydroxide (TCL002)

Foetal bovine serum (RM1112/RM10432)

# **Quality Control:**

# **Appearance**

Off-white to Creamish white, homogenous powder.

#### **Solubility**

Clear solution at 15.5gms/L

#### pH without Sodium Bicarbonate

4.80 - 5.40

## pH with Sodium Bicarbonate

6.60 - 7.20

#### Osmolality without Sodium Bicarbonate

265.00 - 305.00

#### **Osmolality with Sodium Bicarbonate**

310.00 - 350.00

#### **Cultural Response**

The growth promotion capacity of the medium is assessed qualitatively by analyzing the cells for the morphology and quantitatively by estimating the cell counts and comparing it with a control medium through minimum three subcultures.

#### **Endotoxin content**

NMT 5EU/ml

# **Storage and Shelf Life:**

- 1. All the powdered media and prepared liquid culture media should be stored at 2-8°C. Use before the expiry date. Inspite of above recommended storage condition, certain powdered medium may show some signs of deterioration /degradation in certain instances. This can be indicated by change in colour, change in appearance and presence of particulate matter and haziness after dissolution.
- 2. Preparation of concentrated medium is not recommended since free base amino acids and salt complexes having low solubility may precipitate in concentrated medium.
- 3. pH and sodium bicarbonate concentration of the prepared medium are critical factors affecting cell growth. This is also influenced by amount of medium and volume of culture vessel used (surface to volume ratio). For example, in large bottles such as Roux bottles pH tends to rise perceptibly as significant volume of carbon dioxide is released. Therefore, optimal conditions of pH, sodium bicarbonate concentration, surface to volume ratio must be determined for each cell type. We recommend stringent monitoring of pH. If needed, pH can be adjusted by using sterile 1N HCl or 1N NaOH or by bubbling in carbon dioxide.
- 4. If required, supplements can be added to the medium prior to or after filter sterilization observing sterility precautions. Shelf life of the medium will depend on the nature of supplement added to the medium.

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