

SFRE Medium 199-2

**With Earle's Salts, L-Glutamine, Galactose and Glucose
Without Sodium bicarbonate and Insulin**

Product Code: AT090

Product Description :

SFRE Medium 199 is modification of medium 199 developed for growth and maintenance of primary baboon kidney (Bak) cells. Both the media were formulated by supplementing medium M199 with insulin, sodium pyruvate, zinc sulfate, and increasing arginine-HCl, cysteine, cystine, L-glutamine, L-glutamic acid, glycine, histidine, tyrosine, and glucose to maximally active nontoxic concentrations. SFRE 199-2 is additionally supplemented with galactose to avoid excessive accumulation of lactic acid and to maintain pH in the physiological range for prolonged maintenance of the cells.

AT090 is SFRE Medium 199-2 with Earle's salts, L-glutamine, galactose and glucose. It does not contain insulin, hence has to be added separately prior to use. Users are advised to review the literature for recommendations regarding medium supplementation and physiological growth requirements specific for different cell lines.

Composition :

| Ingredients | mg/L |
|---------------------------------------|----------|
| INORGANIC SALTS | |
| Calcium chloride dihydrate | 265.000 |
| Ferric nitrate nonahydrate | 0.720 |
| Magnesium sulphate anhydrous | 97.720 |
| Potassium chloride | 400.000 |
| Sodium acetate | 50.000 |
| Sodium chloride | 6800.000 |
| Sodium dihydrogen phosphate anhydrous | 122.000 |
| Zinc sulphate heptahydrate | 0.100 |
| AMINO ACIDS | |
| Glycine | 100.000 |
| Hydroxy-L-Proline | 10.000 |
| L-Alanine | 25.000 |
| L-Arginine hydrochloride | 150.000 |
| L-Aspartic acid | 30.000 |
| L-Cysteine hydrochloride monohydrate | 4.000 |

| | |
|---------------------------------------|----------|
| L-Cystine dihydrochloride | 43.800 |
| L-Glutamic acid | 75.000 |
| L-Glutamine | 300.000 |
| L-Histidine hydrochloride monohydrate | 40.000 |
| L-Isoleucine | 20.000 |
| L-Leucine | 60.000 |
| L-Lysine hydrochloride | 70.000 |
| L-Methionine | 15.000 |
| L-Phenylalanine | 25.000 |
| L-Proline | 40.000 |
| L-Serine | 25.000 |
| L-Threonine | 30.000 |
| L-Tryptophan | 10.000 |
| L-Tyrosine disodium salt dihydrate | 116.000 |
| L-Valine | 25.000 |
| VITAMINS | |
| Calciferol | 0.100 |
| Choline chloride | 0.500 |
| D-Biotin | 0.010 |
| D-Ca-Pantothenate | 0.010 |
| DL-Tocopherol phosphate disodium salt | 0.010 |
| Folic acid | 0.010 |
| L-Ascorbic acid | 0.050 |
| Menadione sodium bisulphite | 0.016 |
| Niacin | 0.025 |
| Niacinamide | 0.025 |
| Pyridoxal hydrochloride | 0.025 |
| Pyridoxine hydrochloride | 0.025 |
| Retinol Acetate | 0.140 |
| Riboflavin | 0.010 |
| Thiamine hydrochloride | 0.010 |
| i-Inositol | 0.050 |
| p-Amino benzoic acid (PABA) | 0.050 |
| OTHERS | |
| Adenine sulphate | 10.000 |
| Adenosine monophosphate | 1.000 |
| Adenosine triphosphate | 0.200 |
| Cholesterol | 0.200 |
| D(+) Galactose anhydrous | 1000.000 |
| D-Glucose | 2000.000 |
| Deoxyribose | 0.500 |

| | |
|------------------------|---------|
| Glutathione reduced | 0.050 |
| Guanine hydrochloride | 0.300 |
| Hypoxanthine | 0.354 |
| Phenol red sodium salt | 10.000 |
| Polysorbate 80 | 4.900 |
| Ribose | 0.500 |
| Sodium pyruvate | 150.000 |
| Thymine | 0.300 |
| Uracil | 0.300 |
| Xanthine | 0.344 |

Directions :

1. Suspend 12.1gms in 900 ml tissue culture grade water with constant, gentle stirring until the powder is completely dissolved. Do not heat the water.
2. Add 2.20gms of sodium bicarbonate powder (TC230) or 29.3ml of 7.5% sodium bicarbonate solution (TCL013) for 1 litre of medium and stir until dissolved.
3. Adjust the pH to 0.2-0.3 pH units below the desired pH using 1N HCl or 1N NaOH since the pH tends to rise during filtration.
4. Make up the final volume to 1000ml with tissue culture grade water.
5. Sterilize the medium immediately by filtering through a sterile membrane filter with a porosity of 0.22 micron or less, using positive pressure rather than vacuum to minimize the loss of carbon dioxide.
6. Aseptically add sterile supplements as required and dispense the desired amount of sterile medium into sterile containers.
7. Store liquid medium at 2-8°C and in dark till use.

Material required but not provided :

Tissue culture grade water (TCL010)
Sodium bicarbonate (TC230)
Sodium bicarbonate solution, 7.5% (TCL013)
1N Hydrochloric acid (TCL003)
1N Sodium hydroxide (TCL002)
Foetal bovine serum (RM1112/RM10432)

Quality Control:

Appearance

White to light pink, homogenous powder

Solubility

Clear solution at 12.1 gms/L.

pH without Sodium Bicarbonate

4.70 -5.30

pH with Sodium Bicarbonate

7.20 -7.80

Osmolality without Sodium Bicarbonate(mOsm/Kg H₂O)

260.00 -300.00

Osmolality with Sodium Bicarbonate(mOsm/Kg H₂O)

300.00 -340.00

Cultural Response

The growth promotion capacity of the medium is assessed qualitatively by analyzing the cells for the morphology and quantitatively by estimating the cell counts.

Endotoxin Content

NMT 1EU/ml

Storage and Shelf Life:

1. All the powdered media and prepared liquid culture media should be stored at 2-8°C. Use before the expiry date. In spite of above recommended storage condition, certain powdered medium may show some signs of deterioration /degradation in certain instances. This can be indicated by change in colour, change in appearance and presence of particulate matter and haziness after dissolution.
2. Preparation of concentrated medium is not recommended since free base amino acids and salt complexes having low solubility may precipitate in concentrated medium.
3. pH and sodium bicarbonate concentration of the prepared medium are critical factors affecting cell growth. This is also influenced by amount of medium and volume of culture vessel used (surface to volume ratio). For example, in large bottles, such as Roux bottles pH tends to rise perceptibly as significant volume of carbon dioxide is released. Therefore, optimal conditions of pH, sodium bicarbonate concentration, surface to volume ratio must be determined for each cell type. We recommend stringent monitoring of pH. If needed, pH can be adjusted by using sterile 1N HCl or 1N NaOH or by bubbling in carbon dioxide.
4. If required, supplements can be added to the medium prior to or after filter sterilization observing sterility precautions. Shelf life of the medium will depend on the nature of supplement added to the medium.

Disclaimer :

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