

# HiKaryoXL™ RPMI Medium

With L-Glutamine, FBS, Gentamicin sulfate and Sodium bicarbonate

Without PHA-M

1X Liquid Karyotyping Medium

**Product Code: AL173A**

## Intended Use:

HiKaryoXL™ RPMI Medium is a karyotyping medium recommended for short term *in vitro* culture of peripheral blood lymphocytes for cytogenetic studies.

## Principle and Interpretation:

Cytogenetic studies include metaphase and pro-metaphase studies carried out on lymphocytes to detect chromosomal aberrations associated with structural and numerical abnormalities. Lymphocytes come from normal peripheral blood and are mitotically inactive, hence have to be stimulated with a mitogen such as Phytohemagglutinin M (PHA-M) or Phytohemagglutinin P (PHA-P).

AL173A is HiKaryoXL™ RPMI Medium composed of a basal medium RPMI 1640 and supplemented with L-Glutamine, FBS, Gentamicin sulfate and Sodium bicarbonate. Is does not contain PHA-M or PHA-P.

## Composition:

Ingredients	mg/L
<b>INORGANIC SALTS</b>	
Calcium nitrate tetrahydrate	100.000
Magnesium sulphate anhydrous	48.840
Potassium chloride	400.000
Sodium bicarbonate	2000.000
Sodium chloride	6000.000
Sodium phosphate dibasic anhydrous	800.000
<b>AMINO ACIDS</b>	
Glycine	10.000
L-Arginine hydrochloride	241.000
L-Asparagine	50.000
L-Aspartic acid	20.000
L-Cystine dihydrochloride	65.200
L-Glutamic acid	20.000
L-Glutamine	300.000
L-Histidine hydrochloride monohydrate	20.960

L-Hydroxyproline	20.000
L-Isoleucine	50.000
L-Leucine	50.000
L-Lysine hydrochloride	40.000
L-Methionine	15.000
L-Phenylalanine	15.000
L-Proline	20.000
L-Serine	30.000
L-Threonine	20.000
L-Tryptophan	5.000
L-Tyrosine disodium salt	28.830
L-Valine	20.000
<b>VITAMINS</b>	
Choline chloride	3.000
D-Biotin	0.200
D-Ca-Pantothenate	0.250
Folic acid	1.000
Niacinamide	1.000
Pyridoxine hydrochloride	1.000
Riboflavin	0.200
Thiamine hydrochloride	1.000
Vitamin B12	0.005
i-Inositol	35.000
p-Amino benzoic acid (PABA)	1.000
<b>OTHERS</b>	
D-Glucose	2000.000
Fetal Bovine Serum	Proprietary
Glutathione reduced	1.000
Gentamicin sulfate	Proprietary
Phenol red sodium salt	5.300

## Type of Specimen:

Clinical samples – Blood

## Specimen Collection and Handling:

For clinical samples follow appropriate techniques for handling as per established guidelines<sup>1,2</sup>.

1. Disinfect the vacutainer by applying 70% isopropyl alcohol to the rubber stopper.
2. If using blood collection tube containing suitable anticoagulant (EDTA), disinfect the tube by applying 70% isopropyl alcohol.
3. Wait for 1 minute.
4. Palpate vein before disinfection of venipuncture site.
5. Starting at the center, swab the site concentrically with alcohol or chlorhexidine swabs.
6. Allow the disinfectant to dry.  
*Note: Do not palpate the vein at this point without sterile gloves.*
7. Collect the required volume of blood by venipuncture.
8. Mix gently by inverting tube 2 – 3 times to avoid coagulation.
9. Sterilize the needle, syringe and other materials used for blood collection by autoclaving before discarding.

## Warning and Precautions:

In Vitro Diagnostic Use only.

This product should be used by trained healthcare professionals/Lab technicians only.

Not to be used for Self testing.

Read the label before opening the container. Wear protective gloves / protective clothing / eye protection / face protection. Follow proper aseptic techniques while handling specimens and cultures. Standard precautions as per established guidelines should be followed while handling clinical specimens. Safety guidelines may be referred in individual safety datasheets.

## Directions

1. Add PHA-M or PHA-P to HiKaryoXL™ RPMI Medium (AL173A) in required concentration (30 – 50µg/ml).
2. Add freshly collected heparinized whole blood to 10ml of HiKaryoXL™ Medium in T-25cm<sup>2</sup> flasks as per the following recommendations:
  - Normal adults - 0.8ml
  - Infants and children- 0.6ml
  - Women (during pregnancy/ postpartum) - 1.0ml
3. Incubate the flasks at 37°C and 5% CO<sub>2</sub> for 70-72 hours in upright position.
4. To determine optimum incubation time i.e. the peak mitotic index, collect samples at different time intervals between 48-72 hours.  
*Note: Peak mitotic index is most commonly observed at 70-72 hrs.*
5. Add 100µl of 10µg/ml of colchicine and incubate at 37°C and 5% CO<sub>2</sub> for additional 2 hours.  
*Note: Incubation time of less than 1 hour might result in reduced mitotic index.*
6. Transfer the entire content of the flask to a sterile centrifuge tube and centrifuge at 800-1000rpm for 10 minutes.

7. Discard the supernatant and resuspend the pellet in 5ml of warm KCl solution (0.075M) and incubate in a water bath at 37°C for 35 - 40 minutes.  
*Note: Add KCl solution drop wise while agitating the cells.*
8. Add 5ml of freshly prepared ice cold fixative (Acetic acid: methanol, 1:3 parts) and mix gently by inverting.
9. Centrifuge cells at 800-1000rpm for 10min.
10. Discard the supernatant and again add 5ml of freshly prepared ice-cold fixative (acetic acid: methanol, 1:3 parts) with constant mixing. Leave the cells at 4°C for 10-15 min.
11. Centrifuge the cells at 1000rpm for 10 minutes.
12. Repeat step no. 10 and 11.
13. Discard the supernatant and resuspend the pellet in 0.5ml of fresh fixative. Store the tubes in refrigerator at 2 – 8°C until use.
14. Clean the slides with mild detergent and wash thoroughly under tap water to make them grease free.
15. Place the clean slides in a beaker containing water such that they are completely immersed in water. Keep the beaker in a refrigerator at 2 – 8°C and allow the slides to cool.  
*Note: Steps 13 and 14 can be performed during incubation period of 2 hours with colchicine solution to save time.*
16. Mix the cell suspension gently by pipetting up and down. Do not vortex.
17. Tilt the ice cold wet slide at 45° angle and drop 50µl suspension at the bottom of slide with the help of micropipette in such a way that the suspension hits hard on the slide and then runs down surface.
18. Similarly drop 50µl suspension the center and 50µl at the top of the slide.  
*Note: Ensure that the direction of dropping is from bottom to the top.*
19. Allow the slides to air dry. Do not blow.
20. Fix them over a hot plate or a boiling water bath.
21. Stain the slides with required staining solution.

## Materials required but not provided:

HiKaryoXL™ PHA-M Solution (TCL061)  
HiKaryoXL™ PHA-P Solution (TCL071)  
HiKaryoXL™ Colchicine Solution (TCL062) or  
HiKaryoXL™ Colcemid® Solution (TCL074)  
Potassium Chloride solution 0.075M (TCL040)  
Methanol  
Acetic Acid  
Giemsa Stain (TCL083)

## Limitations:

Not applicable.

## Quality control:

### Appearance

Orangish colored, clear solution.

### pH

7.00 - 7.60

### Osmolality in mOsm/Kg H<sub>2</sub>O

340.00 - 380.00

### Sterility

No bacterial or fungal growth is observed after 14 days of incubation, as per USP specification.

### Cultural Response

The growth promotion capacity of the medium is assessed qualitatively by analyzing the cells for the morphology and quantitatively by counting the metaphases.

### Storage and shelf life:

Store at -30°C to -10°C in a freezer that is not self-defrosting. Once thawed, the product is stable for about 30 days at 2 – 8°C. Repeated freezing and thawing reduces mitogenic activity and should be avoided. Once thawed, the medium can be aliquoted into smaller volumes and frozen for future use.

Shelf life is 24 months.

Use before expiry date given on the product label.

### Troubleshooting Tips:

Problem	Cause	Solution
No cell growth or very slow growth	Incubation temperature too high or too low	Check incubator temperature. It should be 37°C ± 0.5°C. Lower temperatures retard the growth rate. Higher temperatures usually result in cell death.
	CO <sub>2</sub> percentage in the incubator too high or too low	Check percentage of CO <sub>2</sub> inside the incubator. It should be 5 ± 0.5%
	Blood used for culture is not fresh	Always use fresh blood
No chromosomes or scattered chromosomes	Cells burst during harvest procedure	Ensure gentle addition of fixative and hypotonic solution
No metaphases	Harvesting not performed in exponential phase	Harvesting should be done between 70 – 72 hours
Chromosomes not well spread or non-uniform	Presence of cell aggregates	Disperse cell clumps before dropping the cell suspension on slide
		Drop the cell suspension on the slide from a height
	Non uniform drying of slide	Avoid blowing and always air dry the slide
	Slides not washed properly and not made grease-free	Ensure that the slides are clean and grease-free
Chromosomes contracted	Prolonged treatment with mitotic inhibitor	Repeat the procedure by treating the culture with mitotic inhibitor for recommended time

### Disposal:

User must ensure safe disposal by autoclaving and / or incineration of used or unusable preparations of this product. Follow established laboratory procedures for disposing infectious materials. The materials that comes into contact with clinical samples must be decontaminated and disposed of in accordance with current laboratory techniques<sup>1, 2</sup>.

### References:

1. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2nd Edition.
2. Jorgensen, J. H., Pfaller, M.A., Carroll, K.C., Funke, G. Landry, M.L., Richter, S.S and Warnock., D.W. (2015), Manual of Clinical Microbiology, 11th Edition. Vol. 1.



In vitro diagnostic medical device



CE Marking



Consult instructions for use



Do not use if package is damaged



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Single use. Not intended to be reprocessed and/or  
used on another patient



Contains biological material of animal origin



Sterilized using aseptic processing techniques



Do not re-use

**Disclaimer :**

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