



Buffered Yeast Agar

M585

Intended Use:

Recommended for cultivation of yeasts and moulds and for the controlling of bottle washing operations in the soft drinks and related industries.

Composition**

| Ingredients | Gms / Litre |
|-------------------------------|-------------|
| Yeast extract | 5.000 |
| Dextrose (Glucose) | 20.000 |
| Ammonium sulphate | 0.720 |
| Ammonium dihydrogen phosphate | 0.260 |
| Agar | 15.000 |
| Final pH (at 25°C) | 5.5±0.2 |

**Formula adjusted, standardized to suit performance parameters

Directions

Suspend 41 grams in 1000 ml purified / distilled water. Heat to boiling to dissolve the medium completely. Sterilize by autoclaving at 115°C for 20 minutes. Mix well and pour into sterile Petri plates.

Principle And Interpretation

Yeasts grow well on a minimal medium containing only dextrose and salts. The addition of yeast extract allows faster growth so that during exponential or log phase growth, the cells divide every 90 minutes (1). Buffered Yeast Agar is prepared as per the modification of the yeast-salt medium described by Davis (2).

The medium contains yeast extract, which supplies B-complex vitamins to stimulate growth. Dextrose is the carbohydrate source. The reaction of this medium can be adjusted to required pH values by the addition of citric or lactic acid to the medium after sterilization. The following table shows the amount of the acids required to be added to 100 ml of Buffered Yeast Agar cooled to 50°C.

Volume of acid to be added to 100 ml of medium to achieve the desired pH

| pH | 1% w/v solution of | 1% w/v solution of |
|------|------------------------------|--------------------|
| | Citric acid monohydrate (ml) | Lactic acid (ml) |
| 4.75 | 1.26 | 0.125 |
| 4.5 | 2.24 | 0.2 |
| 4.25 | 3.92 | 0.3 |
| 4.0 | 6.16 | 0.45 |
| 3.75 | 9.52 | 0.7 |
| 3.5 | 14.56 | 1.17 |

Type of specimen

Bottles used in soft drink industries.

Specimen Collection and Handling

Bunker (3, 4) described a practical method for assessing the efficiency of the bottle cleaning operations. In this method, the bottle under test is converted into a roll-tube culture by coating it internally with the medium. When the agar sets, the bottle is incubated and the colonies are counted and examined. This method gives better results than rinsing the bottle and subsequently plating the rinsings. When used for this purpose, the agar concentration in Buffered Yeast Agar should be increased by 1%w/v (before sterilization).

Warning and Precautions :

Read the label before opening the container. Wear protective gloves/protective clothing/eye protection/ face protection. Follow good microbiological lab practices while handling specimens and culture. Standard precautions as per established guidelines should be followed while handling specimens. Safety guidelines may be referred in individual safety data sheets

Limitations :

This medium is general purpose medium and may not support the growth of fastidious organisms.

Quality Control

Appearance

Cream to yellow homogeneous free flowing powder

Gelling

Firm, comparable with 1.5% Agar gel

Colour and Clarity of prepared medium

Light amber coloured, clear to slightly opalescent gel forms in Petri plates.

Reaction

Reaction of 4.1% w/v aqueous solution at 25°C. pH : 5.5±0.2

pH

5.30-5.70

Cultural Response

Cultural characteristics observed after an incubation at 25-30°C for 48-72 hours.

| Organism | Inoculum (CFU) | Growth | Recovery |
|-------------------------------------------------|-------------------|----------------|----------|
| <i>Candida albicans</i> ATCC 10231 | 50-100 | good-luxuriant | ≥70% |
| <i>Saccharomyces cerevisiae</i> ATCC 9763 | 50-100 | good-luxuriant | ≥70% |
| * <i>Aspergillus brasiliensis</i> ATCC 16404 | 50-100 | good-luxuriant | |

Key *- Formerly known as *Aspergillus niger*

Reference

1. Ausubel, Brent, Kingston, Moore, Seidman, Smith and Struhl, 1994, Current Protocols in Molecular Biology, Current Protocols, Brooklyn, N.Y.
2. Davis J. G., 1931, J. Dairy Res., 3:133.
3. Bunker H. J., 1952, Lab. Prac., 18:354.
4. Bunker H. J., 1956, Wallerstein Lab. Communications, 19(65): 143.

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