



## Lowenstein Jensen Medium Base (L.J. Medium)

M162

### Intended Use:

Recommended for isolation and cultivation of *Mycobacterium* species.

### Composition\*\*

Ingredients	Gms / Litre
L-Asparagine	3.600
Potassium dihydrogen phosphate	2.400
Magnesium sulphate	0.240
Magnesium citrate	0.600
Potato starch, soluble	30.000
Malachite green	0.400

\*\*Formula adjusted, standardized to suit performance parameters

### Directions

Suspend 37.24 grams in 600 ml purified / distilled water containing 12 ml glycerol (for bovine bacteria or other glycerophobic organisms additions of glycerol is not desirable). Heat if necessary to dissolve the medium completely. Sterilize by autoclaving at 15 lbs pressure (121°C) for 15 minutes. Meanwhile prepare 1000 ml of whole egg emulsion collected aseptically. Aseptically add and mix egg emulsion base and Gruft Mycobacterial Supplement (FD053) (if desired) gently to obtain uniform mixture. Distribute in sterile screw capped tubes. Arrange tubes in a slanted position. Coagulate and inspissate the medium in an inspissator water bath or autoclave at 85°C for 45 minutes.

### Principle And Interpretation

Solid media used for isolation and cultivation of Mycobacteria are either egg-based or agar-based. Egg-based media contain whole eggs or egg yolk, potato flour, salts and glycerol and are solidified by inspissation. Of the egg-based media, LowensteinJensen Medium is most commonly used (12). L.J. Medium was originally formulated by Lowenstein, containing congo red and malachite green dyes (10). Jensen (8) modified Lowensteins medium by altering the citrate and phosphate contents, eliminating the congo red dye and by increasing the malachite green concentration. Gruft (5,6) further modified L. J. Medium with the addition of two antimicrobics to increase selectivity. This medium supports the growth of a wide variety of Mycobacteria and can also be used for niacin testing (2).

Penicillin and Nalidixic acid (FD053) along with malachite green prevents growth of the majority of contaminants surviving decontamination of the specimen while encouraging earliest possible growth of *Mycobacteria*. RNA (FD053) acts as stimulant and help to increase the isolation rate of *Mycobacteria*. Do not add glycerol to the medium if bovine or other glycerophobic strains are to be cultured (11). Malachite green serves as an inhibitor and also as pH indicator. Formation of blue zone indicates a decrease in pH by gram-positive contaminants (e.g. *Streptococci* ) and yellow zones of dye destruction by gram-negative bacilli. Proteolytic contaminants cause localized or complete digestion of medium. Hardy et al (1) recommended each specimen to be inoculated and incubated in triplicate,

a. To identify saprophytes at room temperature (25°C).

b. To identify presence or absence of pigmentation by photochromogenes and scotochromogenes at 35°C alternately in light and dark as per the type of organism.

Routinely cultivation is carried out aerobically at 35°C.

### Type of specimen

Clinical samples : Sputum

## Specimen Collection and Handling

For clinical samples: Refer appropriate references for standard test procedures of decontamination and isolation (3,4,7,9,12). After use, contaminated materials must be sterilized by autoclaving before discarding.

## Warning and Precautions

In Vitro diagnostic use only. Read the label before opening the container. Wear protective gloves/protective clothing/eye protection/face protection. Follow good microbiological lab practices while handling specimens and culture. Standard precautions as per established guidelines should be followed while handling clinical specimens. Safety guidelines may be referred in individual safety data sheets.

## Limitations

1. This medium is general purpose medium and may not support the growth of fastidious organisms.
2. Certain gram-positive contaminants (e.g. Streptococci ) and gram-negative bacilli may grow on the medium.
3. Certain Saprophytes may also grow on the medium .
4. Proteolytic contaminants cause localized or complete digestion of medium.

## Performance and Evaluation

Performance of the medium is expected when used as per the direction on the label within the expiry period when stored at recommended temperature.

## Quality Control

### Appearance

Greenish blue to peacock blue homogeneous free flowing powder

### Colour and Clarity of prepared medium

The mixture of sterile basal medium and whole egg emulsion, when inspissated, coagulates to yield pale bluish green coloured, opaque smooth slants

### Cultural Response

Cultural characteristics observed in presence of 5-10% CO<sub>2</sub>, with added egg emulsion base, after an incubation at 35-37°C for 2-4 weeks.

Organism	Growth	Growth with Graft Supplement (FD053)	Colony Characteristic
<i>Mycobacterium avium</i> ATCC 25291	luxuriant	good-luxuriant	smooth, non-pigmented colonies
<i>Mycobacterium gordonae</i> ATCC 14470	luxuriant	good-luxuriant	smooth, yellow, orange colonies
<i>Mycobacterium kansasii</i> ATCC 12478	luxuriant	good-luxuriant	photochromogenic, smooth to rough
<i>Mycobacterium smegmatis</i> ATCC 14468	luxuriant	good-luxuriant	wrinkled, creamy white colonies
<i>M. tuberculosis</i> H37RV ATCC 25618	luxuriant	good-luxuriant	granular, rough, warty, dry friable colonies

## Storage and Shelf Life

Store below 10-30°C in a tightly closed container and the prepared medium at 2-8°C. Use before expiry date on the label. On opening, product should be properly stored dry, after tightly capping the bottle in order to prevent lump formation due to the hygroscopic nature of the product. Improper storage of the product may lead to lump formation. Store in dry ventilated area protected from extremes of temperature and sources of ignition Seal the container tightly after use. Use before expiry date on the label.

Product performance is best if used within stated expiry period.

## Disposal

User must ensure safe disposal by autoclaving and/or incineration of used or unusable preparations of this product. Follow established laboratory procedures in disposing of infectious materials and material that comes into contact with clinical sample must be decontaminated and disposed of in accordance with current laboratory techniques (3,4,7,9).

## Reference

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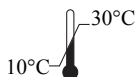
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