



## Tributyryn Agar Base w/o Tributyrin

M157

Tributyryn Agar Base w/o Tributyrin is used for detection of lipolytic microorganisms.

### Composition\*\*

Ingredients	Gms / Litre
Peptic digest of animal tissue	5.000
Yeast extract	3.000
Agar	15.000
Final pH ( at 25°C)	7.5±0.2

\*\*Formula adjusted, standardized to suit performance parameters

### Directions

Suspend 23 grams in 990 ml distilled water. Add 10 ml of Tributyrin (FD081). Mix and heat to boiling to dissolve the medium completely. Sterilize by autoclaving at 15lbs pressure (121°C) for 15 minutes. Shake the flask and individual plate so as to maintain uniform turbidity.

Note : For proper lipase activity, it is recommended to use glass plates instead of disposable plates. Hence USE ONLY GLASS PLATES. DO NOT USE PLASTIC PLATES.

### Principle And Interpretation

Many foods contain significant amount of fats that may be susceptible to hydrolysis. The free fatty acids (FFA) liberated by hydrolysis of fat can be responsible for unpleasant flavours or they may oxidize to compounds with undesirable flavour notes. Many of the problems of fat breakdowns in foods are non-microbial in origin, but numerous bacteria, yeasts and moulds produce lipolytic enzymes that are capable of causing both hydrolytic and oxidative deterioration of fats when present in food samples (1).

Lipolytic enzymatic activities of microorganisms are one of the most important causes for food spoilage and a limited shelf life. Tributyrin Agar was originally formulated by Anderson (2) for the detection and enumeration of lipolytic microorganisms such as *staphylococci* (3), *clostridia* (4), marine *Flavobacteria* and *Pseudomonas* (5) and moulds in foodstuffs and other materials. Tributyrin is the simplest triglyceride occurring in natural fats and oils. It is hydrolyzed by some microorganisms that do not hydrolyze other triglycerides or fats containing longer chain fatty acids. However, for screening purposes, to enumerate lipolytic microorganisms of potential importance in foods, it is the substrate of choice (6, 7).

Peptic digest of animal tissue and yeast extract in the medium provide nutrients to the organisms. Tributyrin degradation by the microorganisms is indicated by clear zones surrounding the lipolytic colonies in the otherwise turbid culture medium. Lipolytic organisms render the medium transparent by converting the fat to water soluble butyric acid (8). The medium should have a uniform turbid emulsion for the effectiveness of the assay (9).

### Quality Control

#### Appearance

Cream to yellow homogeneous free flowing powder

#### Gelling

Firm, comparable with 1.5% Agar gel

#### Colour and Clarity of prepared medium

Light yellow coloured opalescent gel forms with oil droplets in Petri plates.

#### Reaction

Reaction of 2.3% w/v aqueous solution containing 1% Tributyrin at 25°C. pH : 7.5±0.2

#### pH

7.30-7.70

#### Cultural Response

Cultural characteristics observed after an incubation at 35-37°C for 24-48 hours with added Tributyrin (FD081) (under appropriate conditions).

### Cultural Response

Organism	Inoculum (CFU)	Growth	Lipase activity
<b>Cultural Response</b> <i>Clostridium perfringens</i> ATCC 12924	50-100	luxuriant	negative, absence of clear zone around colony
<i>Clostridium sporogenes</i> ATCC 11437	50-100	luxuriant	positive, clear zone around colony
<i>Bacillus subtilis</i> ATCC 6633	50-100	luxuriant	positive, clear zone around colony
<i>Escherichia coli</i> ATCC 25922	50-100	luxuriant	negative, absence of zone around colony
<i>Staphylococcus aureus</i> ATCC 25923	50-100	luxuriant	positive, clear zone around colony

### Storage and Shelf Life

Store below 30°C in tightly closed container and the prepared medium at 2 - 8°C. Use before expiry date on the label.

### Reference

- Vanderzant C. and Splittstoesser D. F., (Eds.), 1992, Compendium of Methods for the Microbiological Examination of Foods, 3rd Ed., APHA, Washington, D.C.
- Anderson J. A., 1939, Ber, IIIrd Int. Mikrobiol. Kongress, 3 : 726
- Innes A. G., 1956, J. Appl. Bacteriol., 19: 39
- Willis A. T., 1960, J. Path. Bacteriol., 80 (2): 379
- Hayes P. R., 1963, J. Gen. Microbiol., 30: 1
- Alford J. A., and Steinle E. E., 1967, J. Appl. Bacteriol., 30: 488
- Frazer T. T., Lawrence R. C., Reiter B., 1967, J. Dairy Sci., 50: 477.
- Collee J. G., Fraser A. G., Marmion B. P., Simmons A., (Eds.), Mackie and McCartney, Practical Medical Microbiology, 1996, 14th Edition, Churchill Livingstone
- MacFaddin J. F., 1985, Media for Isolation-Cultivation-Identification-Maintenance of Medical Bacteria, Vol. 1, Williams and Wilkins, Baltimore

Revision : 02 / 2015

### Disclaimer :

User must ensure suitability of the product(s) in their application prior to use. Products conform solely to the information contained in this and other related HiMedia™ publications. The information contained in this publication is based on our research and development work and is to the best of our knowledge true and accurate. HiMedia™ Laboratories Pvt Ltd reserves the right to make changes to specifications and information related to the products at any time. Products are not intended for human or animal or therapeutic use but for laboratory, diagnostic, research or further manufacturing use only, unless otherwise specified. Statements contained herein should not be considered as a warranty of any kind, expressed or implied, and no liability is accepted for infringement of any patents.