

Technical Data

Yeast Nitrogen Base

M139

Intended Use:

Recommended for classification of yeasts on the basis of their ability to assimilate carbon compounds.

Composition**

Ammonium sulphate 5.000 L-Histidine hydrochloride 0.010 DL-Methionine 0.020 DL-Tryptophan 0.020 Biotin 0.000002 Calcium pantothenate 0.0004 Folic acid 0.00002 Inositol 0.002 Niacin 0.0004 p-Amino benzoic acid (PABA) 0.0002 Pyridoxine hydrochloride 0.0004 Riboflavin (Vitamin B2) 0.0002 Thiamine hydrochloride 0.0004 Porio acid 0.0004	Ingredients	Gms / Litre
DL-Methionine 0.020 DL-Tryptophan 0.020 Biotin 0.000002 Calcium pantothenate 0.0004 Folic acid 0.000002 Inositol 0.002 Niacin 0.0004 p-Amino benzoic acid (PABA) 0.0002 Pyridoxine hydrochloride 0.0004 Riboflavin (Vitamin B2) 0.0002 Thiamine hydrochloride 0.0004	Ammonium sulphate	5.000
DL-Tryptophan 0.020 Biotin 0.000002 Calcium pantothenate 0.0004 Folic acid 0.000002 Inositol 0.002 Niacin 0.0004 p-Amino benzoic acid (PABA) 0.0002 Pyridoxine hydrochloride 0.0004 Riboflavin (Vitamin B2) 0.0002 Thiamine hydrochloride 0.0004	L-Histidine hydrochloride	0.010
Biotin 0.000002 Calcium pantothenate 0.0004 Folic acid 0.000002 Inositol 0.002 Niacin 0.0004 p-Amino benzoic acid (PABA) 0.0002 Pyridoxine hydrochloride 0.0004 Riboflavin (Vitamin B2) 0.0002 Thiamine hydrochloride 0.0004	DL-Methionine	0.020
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Inositol 0.002 Niacin 0.0004 p-Amino benzoic acid (PABA) 0.0002 Pyridoxine hydrochloride 0.0004 Riboflavin (Vitamin B2) 0.0002 Thiamine hydrochloride 0.0004	Calcium pantothenate	0.0004
Niacin 0.0004 p-Amino benzoic acid (PABA) 0.0002 Pyridoxine hydrochloride 0.0004 Riboflavin (Vitamin B2) 0.0002 Thiamine hydrochloride 0.0004	Folic acid	0.000002
p-Amino benzoic acid (PABA) Pyridoxine hydrochloride Riboflavin (Vitamin B2) Thiamine hydrochloride 0.0002 0.0002	Inositol	0.002
Pyridoxine hydrochloride 0.0004 Riboflavin (Vitamin B2) 0.0002 Thiamine hydrochloride 0.0004	Niacin	0.0004
Riboflavin (Vitamin B2) 0.0002 Thiamine hydrochloride 0.0004	p-Amino benzoic acid (PABA)	0.0002
Thiamine hydrochloride 0.0004	Pyridoxine hydrochloride	0.0004
•	Riboflavin (Vitamin B2)	0.0002
Paris acid	Thiamine hydrochloride	0.0004
bone acid	Boric acid	0.0005
Copper sulphate 0.00004	Copper sulphate	0.00004
Potassium iodide 0.0001	Potassium iodide	0.0001
Ferric chloride 0.0002	Ferric chloride	0.0002
Manganese sulphate 0.0004	Manganese sulphate	0.0004
Sodium molybdate 0.0002	Sodium molybdate	0.0002
Zinc sulphate 0.0004	Zinc sulphate	0.0004
Monopotassium phosphate 1.000	Monopotassium phosphate	1.000
Magnesium sulphate 0.500	Magnesium sulphate	0.500
Sodium chloride 0.100	Sodium chloride	0.100
Calcium chloride 0.100	Calcium chloride	0.100
Final pH (at 25°C) 5.4±0.2	Final pH (at 25°C)	5.4±0.2

^{**}Formula adjusted, standardized to suit performance parameters

Directions

For best results, the medium should be prepared in 10X strength. Suspend 6.75 grams in 100 ml purified / distilled water. Add 5 grams of dextrose or an equivalent amount of other carbohydrate. Warm if necessary to dissolve the medium completely. Sterilize by filtration. Keep refrigerated until use. Final medium is made by pipetting 0.5 ml into 4.5 ml of sterile purified / distilled water.

Principle And Interpretation

Yeast Nitrogen Base is formulated as per Wickerham (1) for investigations of yeasts for their different abilities in carbon assimilation. With added carbon source it may also be used for susceptibility testing with antifungal drugs when defined liquid medium is needed (2, 3). Inoculate media tubes with very light inoculum and incubate at 25°C for 6-7 days and again for 20-24 days. Draw lines with India ink on a paper and hold the paper against the Yeast Nitrogen Base tubes. If lines are not seen or appear diffused through the culture, the test is considered positive and if lines are distinguishable, test is negative.

Type of specimen

Pure isolates

Specimen Collection and Handling:

For pure isolates samples follow appropriate techniques for handling specimens as per established guidelines (1,2). After use, contaminated materials must be sterilized by autoclaving before discarding.

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Warning and Precautions:

Read the label before opening the container. Wear protective gloves/protective clothing/eye protection/ face protection. Follow good microbiological lab practices while handling specimens and culture. Standard precautions as per established guidelines should be followed while handling specimens. Safety guidelines may be referred in individual safety data sheets.

Limitations:

1. Use light inoculum Ybeinoculations!

Performance and Evaluation

Performance of the medium is expected when used as per the direction on the label within the expiry period when stored at recommended temperature.

Quality Control

Appearance

Cream to yellow homogeneous free flowing powder

Colour and Clarity of prepared medium

Colourless (at 10X concentration colour of medium is pale yellow) clear solution without any precipitate.

Reaction

Reaction of 0.67% w/v aqueous solution at 25°C. pH: 5.4±0.2

рH

5.20-5.60

Cultural Response

Cultural characteristics observed after an incubation at 25-30°C for 6-7 days (longer if necessary for unto 24 days).

Organism	Growth (Plain)	Growth w. dextrose
Kloeckera apiculata ATCC 9774	none-poor	good
Saccharomyces cerevisiae ATCC 9763 (00058*)	none-poor	good
Saccharomyces uvarum ATCC 28098	none-poor	good

Key: *Corresponding WDCM numbers.

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Disposal

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Reference

- 1. Wickerham, 1951, U.S. Dept. Agri. Tech. Bull No. 1029.
- 2. Lennette E. H., Balows, Hausler and Truant, (Eds.), 1980, Manual of Clinical Microbiology, 3rd Ed., ASM, Washington D.C.
- 3. Padhye A. A., 1981, Diagnostic Procedures for Bacterial, Mycotic and Parasitic Infections, 6th Ed., APHA, Washington, D.C.

Revision: 1/2011

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