



XLD Agar, Modified

M031I

XLD Agar, Modified is recommended for the isolation and enumeration of *Salmonella Typhi* and other *Salmonella* species.

Composition**

Ingredients	Gms / Litre
Yeast extract	3.000
L-Lysine hydrochloride	5.000
Lactose	7.500
Sucrose	7.500
Xylose	3.750
Sodium chloride	5.000
Sodium deoxycholate	1.000
Sodium thiosulphate	6.800
Ferric ammonium citrate	0.800
Phenol red	0.080
Agar	15.000
Final pH (at 25°C)	7.4±0.2

**Formula adjusted, standardized to suit performance parameters

Directions

Suspend 55.43 grams in 1000 ml distilled water. Heat with frequent agitation until the medium boils. DO NOT AUTOCLAVE OR OVERHEAT. Transfer immediately to a water bath at 50°C. After cooling, pour into sterile Petri plates. It is advisable not to prepare large volumes that will require prolonged heating.

Note: Slight precipitation in the medium may occur, which is an inheritant property of the medium, and does not affect the performance of the medium.

Principle And Interpretation

XLD Agar was formulated by Taylor (1-5) for the isolation and differentiation of enteric pathogens including *Salmonella Typhi* from other *Salmonella* species. XLD Agar, Modified (M031I) is recommended for selective isolation and enumeration of *Salmonella Typhi* and other *Salmonella* species in accordance with ISO Committee (20).

XLD Agar has been recommended for the identification of *Enterobacteriaceae* (7) and for the microbiological testing of foods, water and dairy products (8-12). XLD Agar exhibits increased selectivity and sensitivity as compared to other plating media e.g. SS Agar (M108), EMB Agar (M022) and Bismuth Sulphite Agar (M027) (2, 4, 6, and 13-16). The media formulation does not allow the overgrowth of other organisms over *Salmonella* and *Shigella* (17). Samples suspected of containing enteric pathogens, along with other mixed flora, are initially enriched in Modified Semisolid RV Medium Base (M1482) (18).

The medium contains yeast extract, which provides nitrogen and vitamins required for growth. Though the sugars xylose, lactose and sucrose provide sources of fermentable carbohydrates, xylose is mainly incorporated into the medium since it is not fermented by *Shigellae* but practically by all enterics. This helps in the differentiation of *Shigella* species. Sodium chloride maintains the osmotic balance of the medium. Lysine is included to differentiate the *Salmonella* group from the non-pathogens. *Salmonellae* rapidly ferment xylose and exhaust the supply. Subsequently lysine is decarboxylated by the enzyme lysine decarboxylase to form amines with reversion to an alkaline pH that mimics the *Shigella* reaction. However, to prevent this reaction by lysine-positive coliforms, lactose and sucrose are added to produce acid in excess. Degradation of xylose, lactose and sucrose to acid causes phenol red indicator to change its colour to yellow. Bacteria that decarboxylate lysine to cadaverine can be recognized by the appearance of a red colouration around the colonies due to an increase in pH. These reactions can proceed simultaneously or successively, and this may cause the pH indicator to exhibit various shades of colour or it may change its colour from yellow to red on prolonged incubation.

To add to the differentiating ability of the formulation, an H₂S indicator system, consisting of sodium thiosulphate and ferric ammonium citrate, is included for the visualization of hydrogen sulphide produced, resulting in the formation of colonies with

Please refer disclaimer Overleaf.

black centers. The non-pathogenic H₂S producers do not decarboxylase lysine; therefore, the acid reaction produced by them prevents the blackening of the colonies (1).

XLD Agar is both selective and differential medium. It utilizes sodium deoxycholate as the selective agent and therefore it is inhibitory to gram-positive microorganisms. Some *Proteus* strains may give red to yellow colouration with most colonies developing black centers, giving rise to false positive reactions. Non-enterics like *Pseudomonas* and *Providencia* may exhibit red colonies. *S. Paratyphi A*, *S. Choleraesuis*, *S. Pullorum* and *S. Gallinarum* may form red colonies without H₂S, thus resembling *Shigella* species (19).

Quality Control

Appearance

Light yellow to pink homogeneous free flowing powder

Gelling

Firm, comparable with 1.5% Agar gel

Colour and Clarity of prepared medium

Red coloured clear to slightly opalescent gel forms in Petri plates

Reaction

Reaction of 5.54% w/v aqueous solution at 25°C .pH:-7.4±0.2

pH

7.20-7.60

Cultural Response

Cultural response was observed after an incubation at 35-37°C for 18-24 hours. Recovery rate is considered as 100% for bacteria growth on Soyabean Casein Digest Agar.

Cultural Response

Organism	Inoculum (CFU)	Growth	Observed Lot value (CFU)	Recovery	Colour of Colony	Incubation temperature
Cultural Response						
<i>Salmonella Typhimurium</i> ATCC 14028	50 -100	luxuriant	25 -100	≥50 %	red with black centres	18 -72 hrs
<i>Salmonella Abony</i> NCTC 6017	50 -100	good-luxuriant	25 -100	≥50 %	red with black centres	18 -72 hrs
<i>Escherichia coli</i> ATCC 8739	50 -100	fair	10 -30	20 -30 %	yellow	18 -72 hrs
<i>Escherichia coli</i> ATCC 25922	50 -100	fair	10 -30	20 -30 %	yellow	18 -72 hrs
<i>Escherichia coli</i> NCTC 9002	50 -100	fair	10 -30	20 -30 %	yellow	18 -72 hrs
<i>Proteus vulgaris</i> ATCC 13315	50 -100	good-luxuriant	25 -100	≥50 %	grey with black centres	18 -72 hrs
<i>Proteus mirabilis</i> ATCC 25933	50 -100	good-luxuriant		≥50 %	grey with black centres	18 -72 hrs
<i>Salmonella Paratyphi A</i> ATCC 9150	50 -100	good-luxuriant	25 -100	≥50 %	red	18 -72 hrs
<i>Salmonella Paratyphi B</i> ATCC 8759	50 -100	good-luxuriant	25 -100	≥50 %	red with black centres	18 -72 hrs
<i>Salmonella Enteritidis</i> ATCC 13076	50 -100	good-luxuriant	25 -100	≥50 %	red with black centres	18 -72 hrs
<i>Salmonella Typhi</i> ATCC 6539	50 -100	good-luxuriant	25 -100	≥50 %	red with black centres	18 -72 hrs
<i>Shigella dysenteriae</i> ATCC 13313	50 -100	good-luxuriant	25 -100	≥50 %	red	18 -72 hrs
<i>Shigella flexneri</i> ATCC 12002	50 -100	fair-good	15 -40	30 -40 %	red	18 -72 hrs
<i>Shigella sonnei</i> ATCC 25931	50 -100	fair-good	15 -40	30 -40 %	red	18 -72 hrs
<i>Enterobacter aerogenes</i> ATCC 13048	50 -100	fair	10 -30	20 -30 %	yellow	18 -72 hrs
<i>Enterobacter cloacae</i> ATCC 13047	50 -100	fair	10 -30	20 -30 %	yellow	18 -72 hrs
<i>Staphylococcus aureus</i> ATCC 25923	≥10 ³	inhibited	0	0%		≥72 hrs
<i>Staphylococcus aureus</i> ATCC 6538	≥10 ³	inhibited	0	0%		≥72 hrs

Enterococcus faecalis ATCC $\geq 10^3$ 29212 inhibited 0 0% ≥ 72 hrs

Storage and Shelf Life

Store below 30°C in tightly closed container and the prepared medium at 2-8°C. Use before expiry date on the label.

Reference

1. Taylor W. L., 1965, Am. J. Clin. Pathol., 44:471-475.
2. Taylor W. L. and Harris B., 1965, Am. J. Clin. Pathol., 44:476.
3. Taylor W. L. and Harris B., 1967, Am. J. Clin. Pathol., 48:350.
4. Taylor W. L. and Schelhart B., 1967, Am. J. Clin. Pathol., 48:356.
5. Taylor W. L. and Schelhart B., 1968, Am. J. Clin. Pathol., 16:1387.
6. Taylor W. L. and Schelhart B., 1969, Appl. Microbiol., 18:393-395.
7. Chadwick P., Delisle G. H and Byer M., 1974, Can. J. Microbiol., 20, 1653-1664.
8. Downes F. P. and Ito K. (Ed.), 2001, Compendium of Methods for the Microbiological examination of Foods, 4th Ed., APHA Inc. Washington D.C.
9. Eaton A. D., Clesceri L. S., Rice E. W., and Greenberg A. W., (Eds.), 2005, Standard Methods for the Examination of Water and Wastewater, 21st Ed., APHA, Washington, D.C.
10. Wehr H. M. and Frank J. H., 2004, Standard Methods for the Microbiological Examination of Dairy Products, 17th Ed., APHA Inc., Washington, D.C.
11. Williams H., (Ed.), 2005, Official Methods of Analysis of the Association of Official Analytical Chemists, 19th Ed., AOAC, Washington, D.C.
12. FDA Bacteriological Analytical Manual, 2005, 18th Ed., AOAC, Washington, D.C.
13. Dunn C. and Martin W. J., 1971, Appl. Microbiol., 22, 17-22.
14. Rollender M. A., Beckford O., Belsky R. D and Kostroff B. 1969, Am. J. Clin. Pathol., 51, 284-286.
15. Taylor W. L. and Schelhart B., 1969, Appl. Micro. 18, 1387-1392.
16. MacCarthy M. D., 1966, N. Z. J. Med. Lab. Technol., 20, 127-131.
17. Isenberg H. D., Kominos S., and Sigel M., 1969, Appl Microbiol., 18, 656-659.
18. Aspinall S. T., Hindle M. A. and Hutchinson D. N., 1992, 19. Eur. J. Clin. Microbiol., Inf. Dis. 11, 936-939.
- MacFaddin J. F., 1985, Media for Isolation-Cultivation-Identification-Maintenance of Medical Bacteria, Vol. 1, Williams and Wilkins, Baltimore.
20. International Organization for Standardization (ISO), 2002, Draft ISO/DIS 6579:2002.

Revision : 02 / 2015

Disclaimer :

User must ensure suitability of the product(s) in their application prior to use. Products conform solely to the information contained in this and other related HiMedia™ publications. The information contained in this publication is based on our research and development work and is to the best of our knowledge true and accurate. HiMedia™ Laboratories Pvt Ltd reserves the right to make changes to specifications and information related to the products at any time. Products are not intended for human or animal or therapeutic use but for laboratory, diagnostic, research or further manufacturing use only, unless otherwise specified. Statements contained herein should not be considered as a warranty of any kind, expressed or implied, and no liability is accepted for infringement of any patents.