Capsule Stains-Kit

Capsule Stain-Kit is recommended for staining bacterial capsule against dark background.

**Composition**

**Methylene Blue (aqueous)( S021)**

**Ingredients**

- Methylene blue: 0.500gm
- Distilled water: 100.000 ml

**Nigrosin stain, 10% w/v (S025)**

**Ingredients**

- Nigrosin: 10.000 gm
- Formalin: 0.500 ml
- Distilled water: 100.000 ml

**Directions**

For Capsule Staining: Using Nigrosin (S025)

1) To a loopful of cerebrospinal fluid, or to a light aqueous or saline suspension of growth from an agar culture, add a loopful of Nigrosin (S025).
2) Mix well and cover with a thin cover glass. If only a few organisms are present, centrifugation of the cerebrospinal fluid may be necessary.
3) Examine promptly with a high power lens. Light may have to be reduced by lowering the condenser. Oil immersion may be used, if higher magnification is required.

For Capsule Staining: Using Methylene Blue (S021)

1) Transfer aseptically a loopful of culture on a clean and dry slide.
2) Mix it with a loopful of aqueous Methylene Blue (S021).
3) Make a smear by using a glass slide.
4) Allow it to air dry slowly.
5) Observe under oil immersion objective.

**Principle And Interpretation**

Capsules are composed of mucoid polysaccharides of polypeptides. Extracellular capsules are detected by capsule staining. A generally accepted technique for staining capsules employs India ink, nigrosin or congo red (all negative stains) as background material against which the unstained organisms stand out. By counterstaining with dyes like crystal violet or methylene blue, bacterial cell wall takes up the dye. Capsules appear colourless with stained cells against dark background.

**Quality Control**

**Microscopic Examination**

Negative staining is carried out and observed under oil immersion lens.

**Results**

Capsule: Clear halos against dark background

**Storage and Shelf Life**

Store below 30°C in tightly closed container and away from bright light. Use before expiry date on label.

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**Disclaimer:**

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